

FORM PTO-1390
(REV 11-98)

U.S. DEPARTMENT OF COMMERCE PATENT AND TRADEMARK OFFICE

ATTORNEY'S DOCKET NUMBER

**TRANSMITTAL LETTER TO THE UNITED STATES
DESIGNATED/ELECTED OFFICE (DO/EO/US)
CONCERNING A FILING UNDER 35 U.S.C. 371**

TSRI 540.1

U.S. APPLICATION NO. (if known, see 37 CFR 1.5)

09/194889INTERNATIONAL APPLICATION NO.
PCT/US97/09684INTERNATIONAL FILING DATE
4 June 1997PRIORITY DATE CLAIMED
4 June 1996

TITLE OF INVENTION **DIAGNOSTIC AND THERAPEUTIC METHODS RELATED TO REGULATING
ENERGY MOBILIZATION WITH OB PROTEIN AND OB ANTIBODIES**

APPLICANT(S) FOR DO/EO/US **Feng, et al.**

Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:

1. ☒ This is a **FIRST** submission of items concerning a filing under 35 U.S.C. 371.
2. ☐ This is a **SECOND** or **SUBSEQUENT** submission of items concerning a filing under 35 U.S.C. 371.
3. ☒ This express request to begin national examination procedures (35 U.S.C. 371(f)) at any time rather than delay examination until the expiration of the applicable time limit set in 35 U.S.C. 371(b) and PCT Articles 22 and 39(1).
4. ☒ A proper Demand for International Preliminary Examination was made by the 19th month from the earliest claimed priority date.
5. ☒ A copy of the International Application as filed (35 U.S.C. 371(c)(2))
 - a. ☐ is transmitted herewith (required only if not transmitted by the International Bureau).
 - b. ☐ has been transmitted by the International Bureau.
 - c. ☒ is not required, as the application was filed in the United States Receiving Office (RO/US).
6. ☐ A translation of the International Application into English (35 U.S.C. 371(c)(2)).
7. ☒ Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371(c)(3))
 - a. ☐ are transmitted herewith (required only if not transmitted by the International Bureau).
 - b. ☐ have been transmitted by the International Bureau.
 - c. ☐ have not been made; however, the time limit for making such amendments has NOT expired.
 - d. ☒ have not been made and will not be made.
8. ☐ A translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)).
9. ☐ An oath or declaration of the inventor(s) (35 U.S.C. 371(c)(4)).
10. ☐ A translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371(c)(5)).

Items 11. to 16. below concern document(s) or information included:

11. ☐ An Information Disclosure Statement under 37 CFR 1.97 and 1.98.
12. ☐ An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.
13. ☐ A FIRST preliminary amendment.
☐ A SECOND or SUBSEQUENT preliminary amendment.
14. ☐ A substitute specification.
15. ☒ A change of power of attorney and/or address letter.
16. ☒ Other items or information:
 - Return Receipt Postcard
 - Certificate of Express Mailing

U.S. APPLICATION NO. (if known, see 37 CFR 1.5)		INTERNATIONAL APPLICATION NO. PCT/US97/09684		ATTORNEY'S DOCKET NUMBER TSRI 540.1	
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17. ☒ The following fees are submitted:

BASIC NATIONAL FEE (37 CFR 1.492 (a) (1) - (5)) :

Neither international preliminary examination fee (37 CFR 1.482) nor international search fee (37 CFR 1.445(a)(2)) paid to USPTO and International Search Report not prepared by the EPO or JPO **\$970.00**

International preliminary examination fee (37 CFR 1.482) not paid to USPTO but International Search Report prepared by the EPO or JPO. **\$840.00**

International preliminary examination fee (37 CFR 1.482) not paid to USPTO but international search fee (37 CFR 1.445(a)(2)) paid to USPTO **\$760.00**

International preliminary examination fee paid to USPTO (37 CFR 1.482) but all claims did not satisfy provisions of PCT Article 33(1)-(4) **\$670.00**

International preliminary examination fee paid to USPTO (37 CFR 1.482) and all claims satisfied provisions of PCT Article 33(1)-(4) **\$96.00**

ENTER APPROPRIATE BASIC FEE AMOUNT =

Surcharge of **\$130.00** for furnishing the oath or declaration later than ☐ 20 ☐ 30 months from the earliest claimed priority date (37 CFR 1.492(e)).

CLAIMS	NUMBER FILED	NUMBER EXTRA	RATE		
Total claims	35 - 20 =	15	X \$18.00	\$ 270.00	
Independent claims	9 - 3 =	6	X \$78.00	\$ 468.00	
MULTIPLE DEPENDENT CLAIM(S) (if applicable)			+ \$260.00	\$	
TOTAL OF ABOVE CALCULATIONS =				\$ 1408.00	
Reduction of 1/2 for filing by small entity, if applicable. A Small Entity Statement must also be filed (Note 37 CFR 1.9, 1.27, 1.28).				\$	
SUBTOTAL =				\$ 1408.00	
Processing fee of \$130.00 for furnishing the English translation later than <input type="checkbox"/> 20 <input type="checkbox"/> 30 months from the earliest claimed priority date (37 CFR 1.492(f)).				\$	
TOTAL NATIONAL FEE =				\$ 1408.00	
Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31). \$40.00 per property				+	\$
TOTAL FEES ENCLOSED =				\$ 1408.00	
				Amount to be:	\$
				refunded	
				charged	\$

CALCULATIONS **PTO USE ONLY**

a. ☒ A check in the amount of \$ 1,408.00 to cover the above fees is enclosed.

b. ☐ Please charge my Deposit Account No. _____ in the amount of \$ _____ to cover the above fees. A duplicate copy of this sheet is enclosed.

c. ☒ The Commissioner is hereby authorized to charge any additional fees which may be required, or credit any overpayment to Deposit Account No. 19-0962. A duplicate copy of this sheet is enclosed.

NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b)) must be filed and granted to restore the application to pending status.

SEND ALL CORRESPONDENCE TO:

THE SCRIPPS RESEARCH INSTITUTE

Office of Patent Counsel

10550 North Torrey Pines Road, TPC-8

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SIGNATURE:

Thomas Fitting

NAME

34,163

REGISTRATION NUMBER

**DIAGNOSTIC AND THERAPEUTIC METHODS RELATED TO
REGULATING ENERGY MOBILIZATION WITH
OB PROTEIN AND OB ANTIBODIES**

5 Reference to Related Application

This application claims the benefit of U.S. Provisional Application S.N. 60/018,972, filed June 4, 1996, which is incorporated by reference, as are all references cited herein.

10 Governmental Rights

This invention was made with governmental support from the United States Government, National Institutes of Health, Grant DK20043; the United States Government has certain rights in the invention.

15 Background of the Invention

The *obese* gene in human, rat and mouse encodes a protein hormone having an open reading frame 167 amino acid residues in length, called leptin, also known as OB protein or the *ob* gene product. Removal of the signal sequence yields a mature secreted 16 kilodalton protein that is 146 amino acid residues in length.

OB protein is produced primarily by adipocytes of white adipose tissue (WAT). OB protein is secreted directly into the extracellular space and travels through the blood stream. OB protein affects the cells of its target organs by binding to the OB receptor protein, OB-R, that is found on the extracellular surface of the plasma membrane of target cells. Binding of OB protein to OB-R activates the intracellular second messenger cascade of the JAK-STAT system, which is characteristic of activation of cytokine type I receptors.

OB protein is produced in adipocytes in proportion to the mass of stored fat, thereby providing a hormone signal for a lipostatic feedback circuit, which is mediated by the OB receptor. While OB proteins of different species show a close similarity in their sequences, the sequences of OB proteins are not closely similar to other types of proteins. For example, the human *ob* gene

sequence and its mouse homologue (85% sequence identity) have been reported to have no sequence similarity to other proteins of known structure (DiFrancesco, V., et al., Protein Topology Recognition from Secondary Structure Sequences: Application of the Hidden Markov Models to Alpha Class Proteins, J. Mol. Biol. **267**: 446-463 (1997) at page 457).

Although OB protein is composed of a single peptide chain, an intrachain disulfide bond between cysteine 96 and cysteine 146 is required both to stabilize the conformation of the molecule and to confer *in vivo* biological activity (Rock, F.L., et al., The Leptin Haemopoietic Cytokine Fold is Stabilized by an Intrachain Disulfide Bond, Horm. Metab. Res. **28**: 649-652 (1996)). It is believed that the special geometry of the A and D major helices must be maintained in order to dock to a conserved receptor trough in the receptor molecule, a requirement that produces structural similarity between OB proteins and cytokines in the face of negligible sequence conservation (Id. at 651.).

One accepted and successful animal model of human obesity is the genetically obese mouse bearing the recessive *obese* mutation (*ob/ob*). The mouse model reproduces not only the human obesity condition, but also develops non-insulin dependent diabetes mellitus (NIDDM, also known as type II diabetes mellitus). Homologous *obese* genes have been described in mouse, rat and human.

The mouse is also a widely accepted and successful model of sepsis, septic shock and systemic inflammatory response syndrome (SIRS), a term which describes the clinical syndrome of sepsis without regard to its cause. Simple models, involving a large bolus dose of lipopolysaccharide (LPS) administered to mice and using mortality as the primary outcome variable, are well suited for preliminary pharmacological studies of new drugs or other therapeutic agents (Fink, M.P. & Heard, S.O., Laboratory Models of Sepsis and Septic Shock, J. Surg. Res. **49**: 186-196, 1990, at 188-189).

Both the *ob* gene (Zhang, Y., et al., Positional cloning of the mouse *obese* gene and its human homologue, Nature **372**: 425-432 (1994);

accession No. U18812, SEQ ID NO. 1) and its receptor (Tartaglia, L.A., et al., Identification and expression cloning of a Leptin Receptor, Cell **83**: 1263-1271 (1995); Chen, H., et al., Evidence that the Diabetes Gene Encodes the Leptin Receptor: Identification of a Mutation in the Leptin Receptor Gene in *db/db* Mice Cell **84**: 491-495 (1996); accession No. U46135, SEQ ID NO. 2) have been cloned. Shorter versions of the OB receptor, termed the OB-Ra, OB-Rc and OB-Re forms, are produced by alternative splicing of the OB-R mRNA (Lee, G.-H., et al. Nature **379**: 632-635 (1996)). The full length OB receptor is called the OB-Rb form.

Neural activity in specific regions of the central nervous system (CNS), such as the hypothalamus, controls processes related to food intake and energy expenditure. The cloning of the OB protein gene and the OB receptor gene and the localization of OB receptor expression in the hypothalamus has provided supporting evidence for this view as well as suggesting possible mechanisms for relating food intake to stored fat reserves. The OB protein is produced by adipocytes in proportion to the mass of stored fat and, hence, it acts as the signal to a lipostat control circuit. This lipostat signal is transduced at the target cells by the OB receptor, OB-R, in the CNS, resulting changes in neural activity that regulate both food intake and metabolic rate.

Metabolic derangement is an important characteristic of the host response to critical illness called the acute phase response that characterizes conditions such as sepsis and septic shock (Kushner, I. Ann. N.Y. Acad. Sci. **389**: 39-48 (1982)). Hypothermia is a metabolic response that may be pertinent clinical prognostic factor in systemic inflammatory response syndrome in humans (Brivet, F., et al. Crit. Care Med. **22**: 533-534 (1994)).

There is a need for at least one disease marker for systemic inflammatory response syndrome (SIRS) and related conditions. Hereinafter, the term SIRS is used to denote sepsis, septic shock, sepsis syndrome, and related conditions. Disease markers have numerous functions. In this case, a marker for SIRS would be useful for predicting the development of SIRS, identifying patients

with SIRS, predicting outcome, aiding timing and targeting of therapeutic interventions, and determining the pathogenesis of SIRS in patients (Parsons, P.E. & Moss, M. Early Detection and Markers of Sepsis, Clinics in Chest Medicine 17:199-212 (1996)).

5 Summary of the Invention

Embodiments of the present invention include compositions and methods for treating a patient having a condition in which regulating energy metabolism during a systemic inflammatory response is desired, comprising administering a composition having a physiologically effective amount of at least
10 one OB-R agonist ligand. Suitable OB-R agonist ligands include recombinant OB protein, peptide conformational analogs of human OB protein comprising conservative substitutions of amino acid residues and OB-related peptides. A preferred OB-R agonist ligand is recombinant human OB protein.

In another therapeutic embodiment, the beneficial aspects of OB-R
15 agonist ligand administration are facilitated by a coordinated increase in the number of OB receptors (OB-R) produced by the administration of agents that OB-R expression inducers. Suitable OB-R expression inducers include lipopolysaccharide (LPS) and cytokines. Preferred cytokines are interleukin-1 α (IL-1 α), interleukin-1 β (IL-1 β), tumor necrosis factor- α (TNF- α) and interleukin
20 6 (IL-6). Particularly preferred cytokines are IL-6 and IL-1 β .

In a further therapeutic embodiment, antibodies to OB protein are used as agents capable of blocking the effects of OB receptor activation, especially fat mobilization and increased energy utilization. Suitable antibodies to OB proteins may be polyclonal or monoclonal. Suitable antibodies comprise
25 immunoglobulin molecules and immunologically active portions of immunoglobulin molecules, i.e., molecules that contain an antibody combining site or paratope. Exemplary antibody molecules are intact immunoglobulin molecules, substantially intact immunoglobulin molecules and those portions of an immunoglobulin molecule that contain the paratope, including those portions
30 known in the art as Fab, Fab' and F(ab')₂. This embodiment is suitable for

treating metabolic derangement due to conditions such as anorexia, amenorrhea, cachexia and the like.

Antibodies to OB protein are also useful as an assay kit and method for detecting the level of OB protein in a patient. The level of OB protein in a patient is a disease marker that is useful for predicting the development of a condition, identifying patients with the condition, predicting outcome of the condition, aiding timing and targeting of therapeutic interventions, and determining the pathogenesis of the condition in patients. Conditions in which the level of OB protein is a useful marker are SIRS and related conditions such as sepsis and septic shock, as well as anorexia, amenorrhea, cachexia and the like.

Brief Summary of the Drawings

In the drawings:

Figure 1 is a representation of an autoradiogram showing the results of a ribonuclease (RNase) protection assay showing expression of total OB receptor (OB-R_(t)) in lung, kidney and liver at 0, 4, 8 or 24 hours, or 2 (D₂), 3 (D₃) or 5 (D₅) days after intravenous injection of 5 μ g per gram of body weight as well as a dose-response study showing the relative effects on the liver the of injection of 0.05, 0.5 or 5 μ g of LPS per gram of body weight;

Figure 2 is a representation of an autoradiogram showing of the results of a RNase protection assay showing expression of OB-Rb in normal liver (lane 1), and the LPS-treated liver at 24 hours (lane 2), and in the hypothalamus of normal control (lane 3) and *ob/ob* mice (lane 4), compared to OB-R_(m) which represents the mixture of OB-R forms partially protected by the designated nucleotide probes;

Figure 3 is a representation of an autoradiogram showing the results of a RNase protection assay showing expression of OB-Rc in normal liver (lane 1), and the LPS-treated liver at 24 hours (lane 2), and in the hypothalamus of normal control (lane 3) and *ob/ob* mice (lane 4), compared to OB-R_(m) which

represents the mixture of OB-R forms partially protected by the designated nucleotide probes;

Figure 4 is a representation of an autoradiogram showing the results of a RNase protection assay showing expression of OB-Re in normal liver (lane 1), and the LPS-treated liver at 24 hours (lane 2), and in the hypothalamus of normal control (lane 3) and *ob/ob* mice (lane 4), compared to OB-R_(m) which represents the mixture of OB-R forms partially protected by the designated nucleotide probes;

Figure 5 is a representation of an autoradiogram showing the results of a RNase protection assay showing OB-R expression in mouse liver 24 hours after IL-6 (2.5 μ g per mouse), TNF- α (10 μ g per mouse) and IL-1 β (5 μ g per mouse) injection;

Figure 6 is a representation of an autoradiogram showing the results of a RNase protection assay showing OB-R mRNA levels in brain cortex, hypothalamus, and brain stem at various times after LPS injection (5 μ g per gram of body weight);

Figure 7 is a representation of an autoradiogram showing the results of a RNase protection assay showing OB mRNA expression in the adrenal gland (Adr) and white adipose tissue (WAT) at various times after LPS injection (5 μ g per gram of body weight);

Figure 8 is a graphical representation of the construction scheme for the vector pETM1 from a commercially available vector;

Figure 9 is a graphical representation of the weight gain induced by anti-OB antiserum injection in C57BL/6 mice which were given daily injections of anti-OB antiserum (antiOB) or preimmune rabbit sera (control), and whose body weight was measured 12 hours later, where data are expressed as mean \pm standard error of the mean (S.E.M), N=8;

Figure 10 is a graphical representation of the time course of the survival of mice that had received a LPS injection (6 μ g per gram of body weight) after pretreatment with either anti-OB protein antiserum (anti-OB,

N=16), preimmune rabbit serum (control, N=16), or one of three other unrelated rabbit antisera (anti-X, N=9: 3 treated with each antiserum);

Figure 11 is a graphical representation of the time course of the survival of mice that were treated with OB protein (mOB, N=16) or vehicle (control, N=16) after a LPS injection (10 μ g per gram of body weight);

Figure 12 is a graphical representation of the time course of the change in body temperature of mice that had received a LPS injection (6 μ g per gram of body weight) after pretreatment with either anti-OB protein antiserum (anti-OB, N=16, except at * where N<16 due to mortality) or preimmune rabbit serum (control, N=16), data expressed as mean \pm S.E.M.;

Figure 13 is a graphical representation of the time course of the change in body temperature of mice that were treated with OB protein (mOB, N=16) or vehicle (control, N=16, except at * where N<16 due to mortality) after a LPS injection (10 μ g per gram of body weight), data expressed as mean \pm S.E.M.;

Figure 14 is a graphical representation of the time course of the change in body weight (percent of initial body weight) of mice that had received a LPS injection (6 μ g per gram of body weight) after pretreatment with either anti-OB protein antiserum (anti-OB, N=16) or preimmune rabbit serum (control, N=16, except at * where N<16 due to mortality), data expressed as mean \pm S.E.M.;

Figure 15 is a graphical representation of the time course of the change in body weight (percent of initial body weight) of mice that were treated with OB protein (mOB, N=16) or vehicle (control, N=16, except at * where N<16 due to mortality) after a LPS injection (10 μ g per gram of body weight), data expressed as mean \pm S.E.M.; and

Figure 16 is a representation of an autoradiogram showing the results of a RNase protection assay showing the expression of iNOS, IL-1 α , IL-1 β , and TNF- α mRNAs in mouse.

Detailed Description of the Preferred Embodiments

It has been found that substances that initiate or mediate SIRS, for example, LPS and several cytokines, induce the increased expression of OB-R in liver and other peripheral tissues. Thus, occupancy and activation of OB-R by an agonist ligand such as recombinant OB protein, OB-related peptides or peptide conformational analog of human OB protein comprising conservative substitutions of amino acid residues serves as a protective homeostatic mechanism in systemic inflammatory response syndrome conditions such as endotoxic shock, sepsis and septic shock. A preferred OB-R agonist ligand is recombinant OB protein.

Suitable therapeutic human doses of recombinant OB protein are from about 1 micrograms per kilogram body weight to about 50 microgram per kilogram body weight. One preferable therapeutic human dose is about 10 micrograms per kilogram body weight.

While the regulation of energy homeostasis is essentially a function of the CNS, food intake and the majority of the energy expenditure take place in peripheral organs such as the liver. It has been found that the OB protein and the OB receptor have a functional involvement in peripheral energy homeostasis. In general, critical illness and trauma can dramatically alter metabolism, with the expression of the OB receptor changing in response to pathological stress. The expression of OB-R in liver and other peripheral organs, but not in the central nervous system, has now been shown to be induced by endotoxic shock produced intravenous injection of cytokines, such as, IL-1 β , TNF- α and IL-6, as well as cytokine inducing agents such as LPS, into mice, an accepted animal model of SIRS and related conditions. OB protein, antibodies to OB protein, and OB-R expression inducers are useful for the diagnosis and treatment of conditions such as sepsis, systemic inflammatory response syndrome, cachexia and anorexia.

The administration of recombinant mouse OB protein to mice following OB-R induction with a normally lethal dose of LPS conferred complete resistance to LPS, resulting in survival. The OB-treated mice maintained a higher body temperature and displayed dramatic weight loss in contrast to control

counterparts. In vivo administration of OB antisera, on the other hand, elicited the opposite effects by blocking OB-mediated processes, thereby stimulating post-prandial food intake leading to rapid weight gain. Co-administration of LPS with a second in vivo treatment with OB antisera, however, resulted in 100% mortality as compared to animals treated with control antisera.

OB protein, in mediating host responses to LPS-induced endotoxemia, exerts its protective effect primarily by initiating energy mobilization and heat production in critical conditions, the effect of which is proportional to the level of OB protein in the blood. By altering the levels of OB protein, the amount of energy mobilized to resist challenges induced by inflammatory agents is correspondingly altered, thereby effecting the ultimate inflammatory response.

Therefore, in view of the newly discovered physiological properties of OB protein and OB antibodies in regulating energy mobilization and consumption, the present invention describes both diagnostic and therapeutic methods relating to the use of recombinant human OB protein and antibodies thereto in modulating the effects of OB-R activation in vivo.

Diagnostic Applications

OB antibodies are useful for detecting the amount of OB present in sample taken from a patient. One preferred diagnostic embodiment is the use of OB antibodies for detecting the amount of OB present in a blood sample taken from a patient exhibiting a SIRS condition such as sepsis, septic shock, and the like. Such measurements of OB levels in vitro in blood sample by antibody detection is also indicated in various wasting conditions or syndromes associated with several disease states or syndromes, including anorexia, amenorrhea, cachexia, chronic inflammatory conditions, AIDS and AIDS-related conditions, as well as sepsis, septic shock, SIRS and the like. Alternatively, the determination of OB levels is useful in systemic inflammatory response syndromes (SIRS) that are characterized by an acute increase in inflammatory mediators, such as IL-1 β , IL-6, TNF, LPS and the like. Such conditions are noted in preoperative patients

subject to fasting, in patients with acute injuries such as burns or trauma, in patients with SIRS, or with ongoing bacterial infections or those receiving TNF- α for treatment of tumors and in persons suffering from hypothermia.

While there is little sequence similarity between OB proteins and other molecules, the three-dimensional conformation of the OB protein molecule is analogous to that of several long-chain helical cytokines: four major alpha helix regions, A-D, connected by short loops and minor helical regions (Zhang, F., et al., Crystal Structure of the *obese* protein leptin-E100, Nature **387**: 206-209 (1997)).

As used herein, a conformational analog of OB protein is a molecule having substantially the same conformational characteristics of its three-dimensional structure that are required for activation of the OB receptor. Examples of such conformational characteristics include the conformation of the A major helix, the conformation of the D major helix, and the disulfide bond that maintains the geometrical relationship between the A and D major helices. Thus, amino acid substitutions that conserve the conformational characteristics of the molecule, for example, in the loop regions connecting the major helices, would produce conformational analogs to OB protein.

Peptides derived from the region of the OB protein from amino acid residues 106 to 140, as short as about 15 amino acids long, have been shown to be effective in mimicking the action of full-length recombinant OB protein (Grasso, P., et al., In vivo Effects of Leptin-Related Synthetic Peptides on Body Weight and Food Intake in Female *ob/ob* Mice: Localization of Leptin Activity to Domains Between Amino Acid Residues 106 - 140, Endocrinology **138**: 1413-1418 (1997)). As used herein, "OB-related peptides" refers to natural or synthetic peptides derived from the region of the OB protein from about amino acid residue 106 to about amino acid residue 140 and includes conservative amino acid residue substitutions.

The term "antibody" in its various grammatical forms is used herein to refer to immunoglobulin molecules and immunologically active portions

of immunoglobulin molecules, i.e., molecules that contain an antibody combining site or paratope. Exemplary antibody molecules are intact immunoglobulin molecules, substantially intact immunoglobulin molecules and those portions of an immunoglobulin molecule that contain the paratope, including those portions known in the art as Fab, Fab' and F(ab')₂. An antibody composition of the present invention is characterized as containing antibody molecules that immunoreact with OB protein or portions thereof.

An antibody composition of the present invention is typically produced by immunizing a mammal with a inoculum of OB protein or some fragment of OB protein, alone or in combination with a suitable adjuvant such as Freund's adjuvant, and thereby induce in the mammal antibody molecules having the appropriate immunospecificity. The antibody molecules are then collected from the mammal and isolated to the extent desired by well known techniques such as, for example, immunoaffinity chromatography. The antibody composition so produced can be used, inter alia, in the diagnostic methods and systems of the present invention or in the preparation of therapeutic compositions of the present invention.

Monoclonal antibody compositions can also be used with the present invention. A monoclonal antibody composition contains, within detectable limits, only one species of antibody combining site capable of effectively binding to OB protein. Thus, a monoclonal antibody composition of the present invention typically displays a single binding affinity for OB protein even though it may contain antibodies capable of binding proteins other than OB protein. Preferred monoclonal antibodies are those that bind to portions of the OB protein that are required for activation of the OB receptor, such as the A helix, the D helix, or regions of OB protein that maintain the relative positions of the A and D helices that are required for the activation of the OB receptor. Monoclonal antibodies against human OB protein have been described and their preparation was discussed in (Tsuruo, Y. et. al., Horm. Metab. Res. **28**: 753-755 (1996)).

Monoclonal antibodies are also supplied commercially from vendors on a custom

order basis (e.g., Alpha Diagnostic International, Inc., San Antonio, TX).

Purified polyclonal anti-OB antibodies are commercially available from several sources (R&D Systems, Minneapolis, MN; Research Diagnostics, Inc., Flanders, N.J.; Linco Research, Inc., St. Charles, MO; Affinity BioReagents, Inc., Golden, CO).

Determination of OB levels with OB antibodies is performed by assay methods, including ELISA, radioimmunoassay (RIA), Western blot analysis, and the like, that are familiar to one of ordinary skill in the art. The determined OB protein levels are then compared to normal levels for the state of the patient, e.g., fasting, time of day, body mass index (BMI), aerobic conditioning, gender, etc. For example, the normal range found for lean males at 8 a.m. was 12.0 ± 4.4 ng/ml (Sinha, M.K., et al., Nocturnal Rise of Leptin in Lean, Obese, and Non-Insulin-dependent Diabetes Mellitus Subjects, J. Clin. Invest. **97**: 1344-1347 (1996)). See, also Horn, R. et al., Radioimmunoassay for the detection of leptin in human serum, Exp. Clin. Endocrinol. Diabetes **104**: 454-458 (1996); McGregor, G.P., et al., Radioimmunological Measurement of Leptin in Plasma of Obese and Diabetic Human Subjects, Endocrinology **137**: 1501-1504 (1996). It has recently been found that OB protein is present in the circulation in both bound and free form, and that the ratio of the two forms is different in lean and obese subjects (Sinha, M.K., et al., Evidence of Free and Bound Leptin in Human Circulation. Studies in Lean and Obese Subjects and During Short-Term Fasting, J. Clin. Invest. **98**: 1277-1282 (1996)). The relation of free and bound forms to OB protein biological activity can be considered in the context of OB protein assays.

In an alternative embodiment, immunohistochemical assay of OB-R receptor numbers are performed on tissue biopsy materials using standard protocols. A preferred tissue biopsy is liver biopsy.

Therapeutic Applications

Embodiments of the present invention, including methods of administering to a patient compositions comprising OB-R agonist ligands are useful in treating conditions in which it is desirable to regulate or modify energy metabolism during a systemic inflammatory response. Suitable OB-R agonist ligands include recombinant OB protein, peptide conformational analogs of human OB protein comprising conservative substitutions of amino acid residues and OB-related peptides. A preferred OB-R agonist ligand is recombinant human OB protein. A suitable dosage range for recombinant human OB protein is from about 1 microgram per kilogram body weight to about 50 micrograms per kilogram body weight. OB-related peptides are used in a dosage range from about 0.1 microgram per kilogram body weight to about 5 micrograms per kilogram body weight, adjusting the dosage to account for art-recognized differences in potency and solubility (Grasso, P. et al., (1997)).

In a related embodiment, compositions comprising at least one OB-R expression inducer are useful for treating obesity and conditions in which there is an insufficient number of OB receptors, in which low copy number of OB receptors is a limiting factor or in which there is "OB resistance," i.e., a reduced effect of associated with a particular plasma concentration of OB protein.

In general, the OB-R expression inducer is administered in an amount from about 0.003 to about 20 micrograms per kilogram body weight. Suitable OB-R expression inducers include therapeutic cytokines used in cancer therapy, such as IL-1 α , IL-1 β , IL-6 and TNF- α . Suitable dosages and modes of administration are known in the art. For example, a suitable dosage range for IL-1 α is about 0.1 to about 6 micrograms/m²/day. A suitable dosage range for IL-1 β is about 3 to about 200 nanograms/kg/day. A suitable dosage range for IL-6 is about 0.5 to about 20 micrograms/kg/day, with a preferred dosage range for IL-6 being about 1 to about 5 micrograms/kg/day. The therapeutic cytokines may be administered singly or in combination. The mode of administration may be intravenous infusion over an extended time period or a single intravenous or

subcutaneous injection. The daily dose may be administered as a single dose or divided into multiple dose given at intervals during the day.

In a further embodiment, to counteract the possible toxic side effects of OB-R expression inducers such as therapeutic cytokines, such substances are administered in a composition in combination with OB-R agonist ligands. Administration of such compositions is useful for conditions in which cytokines are normally administered for a therapeutic purpose such as tumor treatment, in order to provide effective protection by the OB-R agonist ligands from undesirable metabolic side effects. The up-regulation of the OB-R allows for the complete therapeutic effect mediated by OB-R agonist ligands such as OB protein. A suitable dosage range for recombinant human OB protein is from about 1 microgram per kilogram body weight to about 50 micrograms per kilogram body weight.

In an alternative embodiment, the present invention describes the administration of OB antisera, polyclonal or monoclonal, for treating conditions marked by increased OB and/or increased OB-R activity. Such conditions are various wasting conditions or syndromes associated with several disease states or syndromes, including anorexia, amenorrhea, cachexia, chronic inflammatory conditions, AIDS and AIDS-related conditions. A suitable dosage range for anti-OB protein antibodies is about 0.02 to about 15 milligrams / kg / day.

It has been found that OB protein is an important host defense factor against endotoxin stress. The protective effect of OB protein against endotoxin was not caused by suppressing the expression of major inflammatory mediators, since the mRNA levels of IL-1 α , IL-1 β , TNF- α , and iNOS in lung and spleen were similar in all LPS-treated mice regardless of the experimental manipulations (Figure 16). A comparison of the four groups of mice revealed striking correlations among the OB protein available, the survival from endotoxin shock (Figures 10 and 11), the maintenance of body temperature (Figures 12 and 13), and the loss of body weight (Figures 14 and 15).

Anti-OB Ab-treated mice showed the least loss of body weight and had the most profound hypothermia even with a relatively low dose of LPS. Conversely, OB-treated mice receiving a high dose of LPS maintained a higher body temperature than those control mice that received less LPS. The OB-treated mice also had a greater weight loss than any reported in the literature (16% in the first 24 hours, compared with an average of 10% reported by other groups). The mice in these tests were age, sex and weight matched, fed the same diet, and, therefore, should have had very similar energy store. The different responses to endotoxemia described were likely due to differences in metabolic energy mobilization and dissipation, which, in turn, were attributed to the experimental manipulation of circulating OB protein levels.

When the level of circulating OB protein is varied, the energy mobilized to resist endotoxin challenge is correspondingly altered, and the outcome of the host response to endotoxin stress is affected. *ob/ob* mice, lacking OB protein due to a mutation in the *ob* gene, were very sensitive to LPS insult: a dose as low as 2 μ g per gram of body weight caused a rapid fall of body temperature and death.

The results also suggest the existence of two pathways of thermogenesis and thermostasis. Anti-OB Ab treatment per se did not cause hypothermia in normal mice, suggesting that the thermostasis under non-pathological conditions was largely OB protein-independent. However, when given LPS, the anti-OB Ab-treated mice developed profound hypothermia, indicating that the thermogenesis in response to endotoxemia had become OB protein dependent. A corollary of this model is that genetic defects affecting the OB/OB-R pathway will have a severe hypothermic response to endotoxin. Indeed, *db/db* mice which carry a mutation in the OB-R gene, responded to a low dose of LPS injection in a manner very similar to that seen in *ob/ob* mice (data not shown), despite their increased level of OB expression.

Example 1: Induction of OB-R Expression by Administration of LPS

The injection of LPS and cytokines, substances that are associated with sepsis, septic shock or SIRS caused increases in the expression of the OB receptor in peripheral organs such as liver, but not in brain.

- 5 Intravenous injection of lipopolysaccharide (LPS), IL-1 β , TNF- α and IL-6 to mice induced OB receptor expression in the liver and other peripheral organs, but not in the central nervous system (CNS). To investigate the functional significance of the increased OB-R expression, an anti-OB antiserum was used to neutralize endogenous OB protein in mice prior to an LPS injection.
- 10 The neutralization of OB protein led to profound hypothermia, insignificant loss of body weight, and death in mice in response to an otherwise nonlethal dose of LPS. Conversely, mice administered recombinant mouse OB protein became more resistant to LPS and survived an otherwise lethal dose. The OB protein-treated mice maintained a relatively high body temperature and displayed
- 15 a dramatic weight loss. These results suggest that OB protein may promote energy mobilization to compensate for the increased energy consumption in endotoxemia, and that the OB/OB-R pathway may play an important role in critical host responses to inflammatory stress.

Methods:

- 20 In general, standard techniques or published modifications were used; see, generally, Sambrook, J., et al., Molecular Cloning A Laboratory Manual, 2d Ed., Cold Spring Harbor Laboratory Press (1989). C57BL/6 mice, 5-8 weeks of age and 17-20 g of weight, were injected intravenously with either LPS (5 μ g per gram of body weight, List Biological Laboratory, Campbell, CA),
- 25 IL-1 β (R&D Systems, Minneapolis, MN), IL-6 (2.5 μ g, Pharmingen, San Diego, CA), or TNF- α (a gift from Genentech, San Francisco, CA). Animals were sacrificed at 0, 4, 8 or 24 hours, or 2, 3 or 5 days after the injection.

- Tissue from various organs, including brain, liver and kidney, was dissected and snap-frozen in liquid nitrogen. Total RNA was prepared from the
- 30 frozen tissues by a single-step method (Chomczynski, P. & Sacchi, N., Single-

Step Method of RNA Isolation by Acid Guanidinium Thiocyanate-Phenol-Chloroform Extraction, Analytical Biochemistry **162**: 156-159 (1987).

The RNase protection assays were carried out as previously described (Feng, L., et al., Alternative Splicing of the NC1 Domain of the Human $\alpha 3(\text{IV})$ Collagen Gene J. Biol. Chem. **269**: 2342-2348 (1994); Xia, Y., et al., LPS-Induced MCP-1, IL-1 β , and TNF- α mRNA Expression in Isolated Erythrocyte-Perfused Rat Kidney, Am. J. Physiol. **264**: F774-F780 (1993)). A ten microgram aliquot of total RNA pooled from three similarly treated mice was used for each sample in the RNase protection assay. The stored pooled samples were dissolved in 10 μl of 80% formamide, 0.4 M NaCl, 1 mM EDTA, and 40 mM piperazine-N,N'-bis(2-ethanesulfonic acid), heated to 85 degrees Celsius for 5 minutes. Each ten microgram sample was then hybridized with about 1×10^5 cpm (counts per minute) of the appropriate [^{32}P]UTP-labeled antisense riboprobe at 55 degrees Celsius for at least 10 hours. The unhybridized RNA was then digested with 50 unit/ml RNase T1 (GIBCO/BRL, Gaithersburg, MD) and 24 $\mu\text{g/ml}$ RNase A at 30 degrees Celsius for one hour. The RNase was then digested with 625 $\mu\text{g/ml}$ proteinase K (Boehringer Mannheim, Indianapolis, IN) for 30 minutes at 37 degrees Celsius. After phenol-chloroform extraction and sodium-acetate-ethanol precipitation, the protected hybridized RNA was denatured and electrophoresed on a 10% polyacrylamide gel. The gels were transferred to 3M Whatman filter paper, dried and exposed to Kodak X-Omat film. The resulting autoradiograms were developed in a Kodak X-Omat processor were used only for qualitative screening.

Radioactivity due to hybridization of target sequences with ^{32}P -labeled riboprobes was quantified by scanning the gels on an AMBIS radioanalytic scanning system (AMBIS Systems, San Diego, CA).

An OB-Rb cDNA probe (from base 2548 to base 2835 of OB-Rb, Gen-BankTM Accession No. U46135) was subcloned from a full-length mouse OB-Rb cDNA. The full-length mouse cDNA for the OB-R long form (OB-Rb)

was cloned from a mouse hypothalamus cDNA library (Stratagene, La Jolla, CA), and the sequence was verified against that of U46135.

5 The full-length mouse cDNA for the OB-R short form (OB-Ra) was cloned from a mouse lung cDNA Library (Stratagene, La Jolla, CA). A 224 bp probe that included base 1250 to base 1474 (as indicated on the OB-Rb sequence) of OB-Ra was used for the RNase protection assay. This fragment, which comprises a sequence that is shared by all variants of OB-R, was used as a probe for the total level of OB-R (OBR₀).

10 The expression of other forms of OB-R mRNA was analyzed using selective probes for the respective different forms of OB-R. The designated probes provided full protection to their corresponding OB-R forms and partial protection for other OB-R forms. A probe derived from L32 (33-126, Gen-Bank™ Accession No. X06483), a housekeeping gene encoding ribosomal protein, was used as a control.

15 OB-Rc and OB-Re probes were cloned by reverse-transcription-PCR (RT-PCR) of total liver RNA from LPS-treated C57BL/6 mice. Protocols for RT-PCR are known in the art (for example, pages 15-13 - 15-15 of Ausubel, F.M., et al., Short Protocols in Molecular Biology, 2nd Edition, John Wiley and Sons, New York, (1992)).

20 One suitable protocol for RT-PCR is a modification of that previously described (Feng, L., et al., J. Biol. Chem. **269**: 2342-2348, 1994). The primers used in RT-PCR are listed in Table 1, below. Primer oligonucleotides were synthesized using an ABI model 380B synthesizer (Applied BioSystems, Foster City, Ca).

Table 1: PCR Primers

		Sequence	
	OB-Rc sense	5'-GCTATCGACAAGCAGCAGAAT-3'	(SEQ ID NO. 8)
5	OB-Rc antisense	5'-TGAACACAACAACATAAAGCCC-3'	(SEQ ID NO. 9)
	OB-Re sense	5'-TGTTATATCTGGTTATTATTGAATGG-3'	(SEQ ID NO. 10)
10	OB-Re antisense	5'-CATTAAATGATTTATTATCAGAATTGC-3'	(SEQ ID NO. 11)

First strand cDNA synthesis was performed using total liver RNA from LPS-treated C57BL/6 mice and murine leukemia virus reverse transcriptase with a random hexanucleotide primer. The 100 μ l reaction mixture contained standard enzyme buffer, 5 μ g of total RNA, 20 units of RNasin (RNase inhibitor), 500 pmol of hexanucleotide primer, 10 mM dithiothreitol, 1 mM of each dNTP, with 200 units of reverse transcriptase. Each reaction mixture was heated to 95 degrees Celsius for 10 minutes. PCR was then performed with separate aliquots of the reaction mixture with the appropriate primers for 35 cycles, using 60 degrees Celsius for annealing.

The cDNA segments used to generate riboprobes were excised by the appropriate restriction endonucleases and subcloned into the multiple cloning site of a standard transcription vector. Suitable transcription vectors include a vector chosen from the pGEM series (Promega, Madison, WI). Labelled single stranded riboprobes were synthesized using standard *in vitro* transcription protocols, either those provided by the manufacturer or other standard protocols (e.g. Ausubel, F.M., et al., pages 4-18 - 4-21) with the appropriate (e.g., SP6 or T7) bacteriophage RNA polymerase. The riboprobes contained regions corresponding to the vector polylinker in addition to the region corresponding to the target sequence, and thus were longer than the protected bands. The mouse ribosomal L32 gene, a constitutively expressed "housekeeping" gene, was used throughout the study as a control.

Results:

When LPS was administered to C57BL/6 mice, a strong induction of total OB-R expression (OB-R_(t)) was detected in a number of peripheral organs (Figure 1), but not in several areas of the central nervous system, such as the hypothalamus, that are known to express OB-R (compare Figure 1 and Figure 6). The increased expression of OB-R was most prominent in the liver, the major site of metabolic regulation. The increase of OB-R mRNA expression in the liver was LPS dose-dependent, and peaked between 24 and 48 hours post-LPS injection (Figure 1).

Unexpectedly, RNase protection assays using probes specific for alternatively spliced forms of OB-R mRNA revealed that the long form, OB-Rb, was also induced in the liver to a level comparable to that found in the *ob/ob* mouse hypothalamus and greater than that of the lean control mouse hypothalamus (Figure 2). However, the majority of hepatic OB-R were the OB-Ra, OB-Rc (Figure 3) and OB-Re (Figure 4) forms. OB-Rd expression in the liver was undetectable (data not shown).

In addition to LPS, OB-R expression was induced by the cytokines IL-6, IL-1 α and TNF- α (Figure 5). Contrary to a recent report (12), we found no detectable increase in OB expression in white adipose tissue in LPS-treated mice, but detected a distinct induction of OB mRNA expression in the adrenal gland (Figure 4). No OB mRNA expression was found in the brain, heart, lung, liver, kidney, spleen, muscle, stomach, duodenum, jejunum, ileum, or colon of LPS-treated mice (data not shown).

Example 2: Production of Recombinant OB Protein

Recombinant OB protein was expressed in *E. coli* using a prokaryotic expression vector and extracted from inclusion bodies. Other vectors and host cells systems, including eukaryotic cells, are known in the art and also suitable for the expression of OB protein. See, generally, Ausubel, F.M., et al. Short Protocols in Molecular Biology, 2nd Ed., pages 16-1 to 16-89.

The coding region of mouse OB cDNA (65-619, Gen-Bank™ Accession No.U18812) was cloned by RT-PCR of total RNA from C57BL/6 white adipose tissue. The coding region was subcloned in expression vector, pETM1 (Feng, L., et al., *J. Biol. Chem.* 269: 2342-2348, 1994), to express a His-tagged recombinant mouse OB protein. The construction of pETM1 from the commercially available vector pET-11a (Novagen, Madison, WI) is illustrated on Figure 8.

After the expression of OB protein is induced, the bacteria were harvested and the inclusion bodies were extracted with a buffer containing 6M urea. The extract was loaded on a Ni-NTA affinity column (Qiagen, Chatsworth, CA) and the purification procedure was carried out as previously described (Feng, et al.(1994)). The protein was refolded on the column by adding refolding buffer containing 5 mM CaCl₂/20 mM Tris/0.2 NaCl with an urea gradient of 4 M - 0.5 M at a rate of about 0.5 ml/minute. After refolding, the protein was eluted with 80 mM imidazole/5 mM CaCl₂/20 mM Tris/0.2 NaCl/0.5 mM urea and then dialyzed against phosphate-buffered saline (PBS). Polyclonal antibodies was raised by immunizing a rabbit with the recombinant mouse OB and Freund's adjuvant using standard procedures. Antiserum was used in the following examples.

Example 3: Effects of Anti-OB Antibodies on Metabolism

Intravenous administration of antibodies directed against OB protein effectively opposes the effects of endogenous OB protein.

Recombinant mouse OB protein was produced in an *E. coli* expression system as described in Example 2, and was used to generate rabbit polyclonal anti-OB antibody. The antibody, when injected intravenously into mice, stimulated food intake, leading to rapid weight gain, and thus was effective in blocking OB protein function. The results are shown in Figure 9.

Female C57BL/6 mice, 6-8 weeks of age and 15-17 g of weight, were group housed four per cage and adapted to a 12:12 hour light: dark cycle (light from 6:00 to 18:00). Mice were given a daily intravenous injection of 0.2

ml anti-OB antiserum or preimmune rabbit sera ("vehicle") at 10 p.m., after their initial dark phase food intake. Their body weight was measured at 10 a.m. the next day.

5 The weight gain induced by anti-OB antiserum is illustrated in Figure 9. Data are expressed as mean \pm S.E.M. (N=8). While the weight of the control group remained essentially constant over the week, the anti-OB treated group showed a weight increase at the first weighing, which continued for the entire study period.

10 Example 4: Effects of Anti-OB Antibodies and Recombinant OB-Protein on Response to Endotoxic Shock

The fact that the OB-R variants induced in the liver were predominantly short forms raised the question of functional relevance of the hepatic OB-R expression. OB-Rb is the main form expressed in the hypothalamus, while the choroid plexus expresses only OB-Ra. That the
15 mutation in *db/db* mice affects OB-Rb, but not OB-Ra, suggests that OB-Rb is crucial for regulating food intake and OB-Ra may act as an OB protein transporter. Accordingly, the prominent expression of OB-R in the liver could initiate intracellular signal transduction or, alternatively, mediate the clearance of OB protein. We found that administration of neutralizing anti-OB antibody (Ab)
20 or OB protein to LPS-treated mice distinguished between the two alternatives.

Male C57BL/6 mice, 5-8 weeks of age and 17-21 g of body weight, were used for this study. For antisera treatment, mice were given an i.v. injection of 200 μ l rabbit antisera. LPS at a dose of 6 μ g per gram of body weight was then co-injected with a second dose of anti-OB antibody to the
25 pretreated mice. LPS was dissolved in antisera at a concentration of 0.6 mg/ml and was injected intravenously 4 hours after the initial antisera treatment. Food was retrieved from mouse cages during the 4-hour pretreatment period to prevent any food intake differences resulting from anti-OB antiserum-induced hyperphagia, and was added back after LPS injection.

While this dose of LPS was not lethal to C57BL/6 mice treated with preimmune rabbit serum, all the mice in the anti-OB Ab-treated group died within 40 hours. Figure 10 illustrates the results from three groups of mice: those pretreated with anti-OB antiserum (anti-OB, N=16), preimmune rabbit sera control, N=16), or three other unrelated rabbit antisera ("Anti-X", N=3 for each antiserum). This LPS sensitizing effect was specific for anti-OB Ab, since mice treated similarly with three other unrelated antibodies ("Anti-X") all survived (Figure 10).

In comparison, mice treated with OB protein (5 μ g per gram of body weight mOB, N=16) were able to survive a higher dose of LPS (10 μ g per gram of body weight) that was fatal to the control group of mice receiving the vehicle alone (control, N=16). Mouse OB protein and LPS were prepared in saline at a concentration of 0.5 mg/ml and 1 mg/ml, respectively, and injected intravenously into the mice. A vehicle solution used for OB protein dialysis was injected into control mice. To eliminate any circadian effect, experiments at different days were all started at the same hours. Mice were examined at 4-hour intervals post LPS injection for the first 24 hours, and survival was monitored for 7 days following the LPS injection. The results are shown in Figure 11. The dose of LPS killed all the mice in the control group within 24 hours. However, in the experimental group, OB protein treatment conferred mice complete resistance to this dose of endotoxin. The OB-treated mice displayed noticeably less severe symptoms of endotoxemia, remaining alert and responsive to touch and other manipulation, and recovering quickly.

The effects of both OB protein and anti-OB antibodies on body temperature and body weight were monitored in the same groups of mice. Core body temperature measurements were made by insertion of a thermistor probe (Yellow Springs Instrument, Yellow Springs, Ohio) into the colon, 1.5 cm beyond the rectum. Body weight measurements were made on a portable digital balance (Ohaus, Florham Park, NJ).

The mice receiving anti-OB antibodies, which ultimately died, showed lower body temperature (Figure 12) and less weight loss (Figure 14) than the corresponding control group than survived. Conversely, the mice receiving OB-protein, and which survived the endotoxic shock, showed higher body temperature (Figure 13) and more weight loss (Figure 15) than the corresponding control group which succumbed.

While administration of both OB protein and anti-OB antibodies had significant effects on survival, body weight and body temperature, there was little effect on the expression of iNOS, IL-1 α , IL-1 β , and TNF- α mRNAs in lung and spleen (Figure 16). Mice were treated as described in above (Figures 10-15).

RNase protection assays were performed as described in Example 1. Each sample was 5 μ g total RNA was used for each sample in the RNase protection assay. Riboprobes were produced as described in Example 1 based on the following cDNA fragments: IL-1 α (from base 172 to base 366, Accession No. X01450, SEQ ID NO. 3), IL-1 β (from base 500 to base 671, Accession No. M15131, SEQ ID NO. 4), TNF- α (from base 428 to base 557, Accession No. M11731, SEQ ID NO. 5), mouse iNOS (from base 2404 to base 2698, Accession No. M92649, SEQ ID NO. 6), and L32 (from base 33 to base 126, Accession No. X064383, SEQ ID NO. 7).

Relatively little change in the pattern of expression of these markers was observed (Figure 16), suggesting that these the protective actions of OB proteins are direct and not indirect and mediated by these cytokines.

The foregoing is intended to be illustrative of the present invention, but not limiting. Numerous variations and modifications of the present invention may be effected without departing from the true spirit and scope of the invention.

SEQUENCE LISTING

(1) GENERAL INFORMATION:

- (i) APPLICANT: Feng, Lili
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Xia, Yiyang
- (ii) TITLE OF INVENTION: DIAGNOSTIC AND THERAPEUTIC METHODS
RELATED TO REGULATING ENERGY MOBILIZATION WITH OB PROTEIN
AND OB ANTIBODIES
- (iii) NUMBER OF SEQUENCES: 11
- (iv) CORRESPONDENCE ADDRESS:
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 - (C) CITY: Chicago
 - (D) STATE: IL
 - (E) COUNTRY: US
 - (F) ZIP: 60606
- (v) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Floppy disk
 - (B) COMPUTER: IBM PC compatible
 - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 - (D) SOFTWARE: PatentIn Release #1.0, Version #1.30
- (vi) CURRENT APPLICATION DATA:
 - (A) APPLICATION NUMBER:
 - (B) FILING DATE: 04-JUN-1997
 - (C) CLASSIFICATION:
- (vii) PRIOR APPLICATION DATA:
 - (A) APPLICATION NUMBER: US 60/018,972
 - (B) FILING DATE: 04-JUN-1996
- (viii) ATTORNEY/AGENT INFORMATION:
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 - (B) REGISTRATION NUMBER: 30,203
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 - (B) TELEFAX: 312-580-1189

(2) INFORMATION FOR SEQ ID NO:1:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 2793 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:

(A) ORGANISM: *Mus musculus*

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

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TGCCTATCCA GAAAGTCCAG GATGACACCA AAACCTCAT CAAGACCATT GTCACCAGGA 180
TCAATGACAT TTCACACACG CAGTCGGTAT CCGCCAAGCA GAGGGTCACT GGCTTGGACT 240
TCATTCTTGG GCTTCACCCC ATTCTGAGTT TGTCCAAGAT GGACCAGACT CTGGCAGTCT 300
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AGAATCTCCG AGACCTCCTC CATCTGCTGG CTTTCTCCAA GAGCTGCTCC CTGCCTCAGA 420
CCAGTGGCCT GCAGAAGCCA GAGAGCCTGG ATGGCGTCCT GGAAGCCTCA CTCTACTCCA 480
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GCGCTCAAGA TTGACCTCTG GTGACTGGTT TTGTTTCTAT TGTGACTGAC TCTATCCAAA 1260
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 GAGTTGGAGG TAGATTTTGG AGGATCTGAG GGC 2793

(2) INFORMATION FOR SEQ ID NO:2:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 3862 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (vi) ORIGINAL SOURCE:
 - (A) ORGANISM: Mus musculus

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

GTCGACCCAC GCGTCCGGAG GAATCGTTCT GCAAATCCAG GTGTACACCT CTGAAGAAAG 60
 ATGATGTGTC AGAAATTCTA TGTGGTTTTG TTACACTGGG AATTTCTTTA TGTGATAGCT 120

GCACTTAACC TGGCATATCC AATCTCTCCC TGGAAATTTA AGTTGTTTTG TGGACCACCG	180
AACACAACCG ATGACTCCTT TCTCTCACCT GCTGGAGCCC CAAACAATGC CTCGGCTTTG	240
AAGGGGGCTT CTGAAGCAAT TGTTGAAGCT AAATTTAATT CAAGTGGTAT CTACGTTTCCT	300
GAGTTATCCA AAACAGTCTT CCACTGTTGC TTTGGGAATG AGCAAGGTCA AAAGTGTCTT	360
GCACTCACAG ACAACACTGA AGGGAAGACA CTGGCTTCAG TAGTGAAGGC TTCAGTTTTT	420
CGCCAGCTAG GTGTAAACTG GGACATAGAG TGCTGGATGA AAGGGGACTT GACATTATTC	480
ATCTGTCATA TGGAGCCATT ACCTAAGAAC CCCTTCAAGA ATTATGACTC TAAGGTCCAT	540
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TTTCAGACTG TCCAATGCAA CTGCAGTCTT CGGGGATGTG AATGTCATGT GCCGGTACCC	660
AGAGCCAAAC TCAACTACGC TCTTCTGATG TATTTGGAAA TCACATCTGC CGGTGTGAGT	720
TTTCAGTCAC CTCTGATGTC ACTGCAGCCC ATGCTTGTTG TGAAACCCGA TCCACCCTTA	780
GGTTTGCATA TGGAAGTCAC AGATGATGGT AATTTAAAGA TTTCTTGGA CAGCCAAACA	840
ATGGCACCAT TTCCGCTTCA ATATCAGGTG AAATATTTAG AGAATTCTAC AATTGTAAGA	900
GAGGCTGCTG AAATTGTCTC AGCTACATCT CTGCTGGTAG ACAGTGTGCT TCCTGGATCT	960
TCATATGAGG TCCAGGTGAG GAGCAAGAGA CTGGATGGTT CAGGAGTCTG GAGTGAAGTG	1020
AGTTCACCTC AAGTCTTTAC CACACAAGAT GTTGTGTATT TTCCACCCAA AATTCTGACT	1080
AGTGTGAGGAT CGAATGCTTC TTTTCATTGC ATCTACAAAA ACGAAAACCA GATTATCTCC	1140
TCAAAACAGA TAGTTTGGTG GAGGAATCTA GCTGAGAAAA TCCCTGAGAT ACAGTACAGC	1200
ATTGTGAGTG ACCGAGTTAG CAAAGTTACC TTCTCCAACC TGAAAGCCAC CAGACCTCGA	1260
GGGAAGTTTA CCTATGACGC AGTGTACTGC TGCAATGAGC AGGCGTGCCA TCACCGCTAT	1320
GCTGAATTAT ACGTGATCGA TGTCAATATC AATATATCAT GTGAAACTGA CGGGTACTTA	1380
ACTAAAATGA CTTGCAGATG GTCACCCAGC ACAATCCAAT CACTAGTGGG AAGCACTGTG	1440
CAGCTGAGGT ATCACAGGCG CAGCCTGTAT TGTCCTGATA GTCCATCTAT TCATCCTACG	1500
TCTGAGCCCA AAAACTGCGT CTTACAGAGA GACGGCTTTT ATGAATGTGT TTTCCAGCCA	1560
ATCTTTCTAT TATCTGGCTA TACAATGTGG ATCAGGATCA ACCATTCTTT AGGTTCACCT	1620
GACTCGCCAC CAACGTGTGT CCTTCCTGAC TCCGTAGTAA AACCCTACC TCCATCTAAC	1680
GTAAAAGCAG AGATTACTGT AAACACTGGA TTATTGAAAG TATCTTGGA AAAGCCAGTC	1740
TTTCCGGAGA ATAACCTTCA ATTCCAGATT CGATATGGCT TAAGTGGAAA AGAAATACAA	1800
TGGAAGACAC ATGAGGTATT CGATGCAAAG TCAAAGTCTG CCAGCCTGCT GGTGTCAGAC	1860
CTCTGTGCAG TCTATGTGGT CCAGGTTTCG TGCCGGCGGT TGGATGGACT AGGATATTGG	1920
AGTAATTGGA GCAGTCCAGC CTATACGCTT GTCATGGATG TAAAAGTTCC TATGAGAGGG	1980

662200 662200 662200

CCTGAATTTT GGAGAAAAAT GGATGGGGAC GTTACTAAAA AGGAGAGAAA TGTCACCTTG 2040
CTTTGGAAGC CCCTGACGAA AAATGACTCA CTGTGTAGTG TGAGGAGGTA CGTGGTGAAG 2100
CATCGTACTG CCCACAATGG GACGTGGTCA GAAGATGTGG GAAATCGGAC CAATCTCACT 2160
TTCCTGTGGA CAGAACCAGC GCACACTGTT ACAGTTCTGG CTGTCAATTC CCTCGGCGCT 2220
TCCCTTGTGA ATTTTAACCT TACCTTCTCA TGGCCCATGA GTAAAGTGAG TGCTGTGGAG 2280
TCACTCAGTG CTTATCCCCT GAGCAGCAGC TGTGTCATCC TTTCCTGGAC ACTGTCACCT 2340
GATGATTATA GTCTGTTATA TCTGGTTATT GAATGGAAGA TCCTTAATGA AGATGATGGA 2400
ATGAAGTGGC TTAGAATTCC CTCGAATGTT AAAAAGTTTT ATATCCACGA TAATTTTATT 2460
CCCATCGAGA AATATCAGTT TAGTCTTTAC CCAGTATTTA TGGAAGGAGT TGGAAAACCA 2520
AAGATAATTA ATGGTTTTCAC CAAAGATGCT ATCGACAAGC AGCAGAATGA CGCAGGGCTG 2580
TATGTCATTG TACCCATAAT TATTTCTCTT TGTGTCCTAC TGCTCGGAAC ACTGTTAATT 2640
TCACACCAGA GAATGAAAAA GTTGTTTTGG GACGATGTTC CAAACCCCAA GAATTGTTCC 2700
TGGGCACAAG GACTGAATTT CCAAAGCCT GAAACATTTG AGCATCTTTT TACCAAGCAT 2760
GCAGAATCAG TGATATTTGG TCCTCTTCTT CTGGAGCCTG AACCATTTC AGAAGAAATC 2820
AGTGTGATA CAGCTTGAA AAATAAAGAT GAGATGGTCC CAGCAGCTAT GGTCTCCCTT 2880
CTTTTGACCA CACCAGACCC TGAAAGCAGT TCTATTTGTA TTAGTGACCA GTGTAACAGT 2940
GCTAACTTCT CTGGGTCTCA GAGCACCCAG GTAACCTGTG AGGATGAGTG TCAGAGACAA 3000
CCCTCAGTTA AATATGCAAC TCTGGTCAGC AACGATAAAC TAGTGGAAC TGATGAAGAG 3060
CAAGGGTTTA TCCATAGTCC TGTGAGCAAC TGCATCTCCA GTAATCATTC CCCACTGAGG 3120
CAGTCTTTCT CTAGCAGCTC CTGGGAGACA GAGGCCAGA CATTTTTCCT TTTATCAGAC 3180
CAGCAACCCA CCATGATTTT ACCACAACCT TCATTCTCGG GGTGGATGA GCTTTTGGAA 3240
CTGGAGGGAA GTTTTCCTGA AGAAAATCAC AGGGAGAAGT CTGTCTGTTA TCTAGGAGTC 3300
ACCTCCGTCA ACAGAAGAGA GAGTGGTGTG CTTTGTACTG GTGAGGCAGG AATCCTGTGC 3360
ACATTCCCAG CCCAGTGTCT GTTCAGTGAC ATCAGGATCC TCCAGGAGAG ATGCTCACAC 3420
TTTGTAGAAA ATAATTTGAG TTTAGGGACC TCTGGTGAGA ACTTTGTACC TTACATGCCC 3480
CAATTTCAA CCGTGTCCAC GCACAGTCAC AAGATAATGG AGAATAAGAT GTGTGACTTA 3540
ACTGTGTAAT CTCATCCAAG AAGCCTCAAG GTTCCATTCC AGTAGAGCCT GTCATGTATA 3600
ATGTGTTCTT TTATTGTTGT GGATGTGGGA GACAAGTGTC AGAATCTAGT GTGAAAATGA 3660
TTGTTTCCAA ACTAAGTGTG TCTATTTTCT CTCAGTAATA CAATGAAACA TATGAGGAAG 3720
CCCTCATTA TCTAGTAATG TAGATGGACT CTTACTGAAT ATATTCCCAA GATACTTGGG 3780
GAAGTCTCCC TAATTCTAGC TAAAAATAA CCCAGGAATA GAACTACTAA AACTGAATC 3840

662260 664670

TGGAAAAAAAA AAAAAAAAAA AG

3862

(2) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1974 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:

(A) ORGANISM: Mus musculus

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

AAGTCTCCAG GGCAGAGAGG GAGTCAACTC ATTGGCGCTT GAGTCGGCAA AGAAATCAAG	60
ATGGCCAAAG TTCCTGACTT GTTTGAAGAC CTAAAGAACT GTTACAGTGA AAACGAAGAC	120
TACAGTTCTG CCATTGACCA TCTCTCTCTG AATCAGAAAT CCTTCTATGA TGCAAGCTAT	180
GGCTCACTTC ATGAGACTTG CACAGATCAG TTTGTATCTC TGAGAACCTC TGAAACGTCA	240
AAGATGTCCA ACTTCACCTT CAAGGAGAGC CGGGTGACAG TATCAGCAAC GTCAAGCAAC	300
GGGAAGATTC TGAAGAAGAG ACGGCTGAGT TTCAGTGAGA CCTTCACTGA AGATGACCTG	360
CAGTCCATAA CCCATGATCT GGAAGAGACC ATCCAACCCA GATCAGCACC TTACACCTAC	420
CAGAGTGATT TGAGATACAA ACTGATGAAG CTCGTCAGGC AGAAGTTTGT CATGAATGAT	480
TCCCTCAACC AAATATATA TCAGGATGTG GACAAACACT ATCTCAGCAC CACTTGGTTA	540
AATGACCTGC AACAGGAAGT AAAATTTGAC ATGTATGCCT ACTCGTCGGG AGGAGACGAC	600
TCTAAATATC CTGTTACTCT AAAAATCTCA GATTCACAAC TGTTCTGTGAG CGCTCAAGGA	660
GAAGACCAGC CCGTGTTGCT GAAGGAGTTG CCAGAAACAC CAAAACATCAT CACAGGTAGT	720
GAGACCGACC TCATTTTCTT CTGGAAAAGT ATCAACTCTA AGAACTACTT CACATCAGCT	780
GCTTATCCAG AGCTGTTTAT TGCCACCAA GAACAAAGTC GGGTGACACT GGCACGGGGA	840
CTGCCCTCTA TGACAGACTT CCAGATATCA TAAAAGCAGC CTTATTTCTG GAGTCTATTC	900
ACTTGGGAAG TGCTGACAGT CTGTATGTAC CATGTACAGG AACCTTCCTC ACCCTGAGTC	960
ACTTGACACAG CATGTGCTGA GTCTCTGTAA TTCTAAATGA ATGTTTACCC TCTTTGTAAG	1020
AGAAGAGCAA ACCCTAGTGG AGCCACCCCG ACATATGATA CTATCTGTTA TTTTAAAGAG	1080
TACCCTATAG TTTGCTCAGT ACTAATCATT TTAATTACTA TTCTGCATGG CATTCTTAGG	1140
AGGATCAAAA AGACTCTACA CATATTACAG ATGGGTTAAC AAAGGGATAA AACAACTGAA	1200

AAGCACACTC	AATGCATTTG	GAATATAAAT	TCACAGACCA	ATCTCACTGT	GCACCTTCGG	1260
CTTCAAAATG	CCAGTTGAGT	AGGATAAAGG	TATAAGAACT	TAATGCTGTC	ATTTTCAAAA	1320
GGAAGGGGAC	AATAGCTACA	TCTTTCTTAC	CTCAGTGGGT	TTTACTCCAG	TGAGATCATT	1380
TGGATGAAAT	CCTCCTGTAA	CAGACCTCAA	GAAGGAGACA	GACTGTTGAA	TGTTATTTTT	1440
AAGTTATTTT	ATATATGTAT	TTATAAATAT	ATTTATGATA	ATTATATTAT	TTATGGAACA	1500
TCCTTAAATC	CTCTGAGCTT	GACAGGCATC	CTCACAGCAG	GATTTTCTAG	GTGGTCAGTT	1560
AGATATAGTT	TCCTCTAGAG	CACCATGCTA	CAGACTTTAC	ACTTTTTCCA	CAGCCACGAA	1620
GCTCTCTGTA	CATTCTGTGA	CTTGGGAGCC	CTTTCATCAT	GATCTTAATC	TGTACTGTTT	1680
ACTTTGTTCA	TCTAAAATGA	TAATTGAGTC	AGTCTTTTTT	CCTCCCATCC	TTAAAGCTGT	1740
CTGGGTATTC	TTACATCATT	CAGTCTCACC	TGTAACAAAC	ACCAACCATC	TAAAGATGGA	1800
AAGAGCTTAA	CTGTGACAAC	CACATCACTG	TTACCTGAAG	TTTCTTTTCT	AGAATGTAAT	1860
CAGTGTTTCC	CCTGGATTCC	AATTTTTTTT	TCAAACCACA	GTATCATGTA	ACTATCAACA	1920
ATAACAATCA	ACTCATTATT	ATTAATCATA	ATTAAATAAA	ACAAGTTTGA	GCTG	1974

(2) INFORMATION FOR SEQ ID NO:4:

- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 1339 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:
(A) ORGANISM: *Mus musculus*

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

TGCAGGGTTC	GAGGCCTAAT	AGGCTCATCT	GGGATCCTCT	CCAGCCAAAGC	TTCTTTGTGC	60
AAGTGTCTGA	AGCAGCTATG	GCAACTGTTC	CTGAACTCAA	CTGTGAAATG	CCACCTTTTG	120
ACAGTGATGA	GAATGACCTG	TTCTTTGAAG	TTGACGGACC	CCAAAAGATG	AAGGGCTGCT	180
TCCAAACCTT	TGACCTGGGC	TGTCCAGATG	AGAGCATCCA	GCTTCAAATC	TCACAGCAGC	240
ACATCAACAA	GAGCTTCAGG	CAGGCAGTAT	CACTCATTGT	GGCTGTGGAG	AAGCTGTGGC	300
AGCTACCTGT	GTCTTTCCCG	TGGACCTTCC	AGGATGAGGA	CATGAGCACC	TTCTTTTCCT	360
TCATCTTTGA	AGAAGAGCCC	ATCCTCTGTG	ACTCATGGGA	TGATGATGAT	AACCTGCTGG	420
TGTGTGACGT	TCCCATTAGA	CAGCTGCACT	ACAGGCTCCG	AGATGAACAA	CAAAAAAGCC	480

TCGTGCTGTC GGACCCATAT GAGCTGAAAG CTCTCCACCT CAATGGACAG AATATCAACC 540
 AACAAAGTGAT ATTCTCCATG AGCTTTGTAC AAGGAGAACC AAGCAACGAC AAAATACCTG 600
 TGGCCTTGGG CCTCAAAGGA AAGAATCTAT ACCTGTCCCTG TGTAATGAAA GACGGCACAC 660
 CCACCCTGCA GCTGGAGAGT GTGGATCCCA AGCAATACCC AAAGAAGAAG ATGGAAAAGC 720
 GGTTCGTCTT CAACAAGATA GAAGTCAAGA GCAAAGTGGA GTTTGAGTCT GCAGAGTTCC 780
 CCAACTGGTA CATCAGCACC TCACAAGCAG AGCACAAGCC TGTCTTCCTG GGAAACAACA 840
 GTGGTCAGGA CATAATTGAC TTCACCATGG AATCTGTGTC TTCCTAAAGT ATGGGCTGGA 900
 CTGTTTCTAA TGCCTTCCCC AGGGCATGTG AAGGAGCTCC CTTGTCATGA ATGAGCAGAC 960
 AGCTCAATCT CTAGGACACT CCTTAGTCCT CGGCCAAGAC AGGTCGCTCA GGGTCACAAG 1020
 AAACCATGGC ACATTCTGTT CAAAGAGAGC CTGTGTTTCC TCCTTGCCTC TGATGGGCAA 1080
 CCACTTACCT ATTTATTTAT GTATTTATTG ATTGGTTGAT CTATTTAAGT TGATTCAAGG 1140
 GGACATTAGG CAGCACTCTC TAGAACAGAA CCTAGCTGTC AACGTGTGGG GGATGAATTG 1200
 GTCATAGCCT TGCACTTGAG GTCTTTCATT GAAGCTGAGA ATAAATAGGT TCCTATAATA 1260
 TGGATGAGAA TTTTATGAA TGAAGCATTG GCACATTGCT TTGATGAGTA TGAAATAAAT 1320
 TTCATTAAAC AAACAAACA 1339

(2) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1629 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: Mus musculus

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

GCTGAGGGAC TAGCCAGGAG GGAGAACAGA AACTCCAGAA CATCCTGGAA ATAGCTCCCA 60
 GAAAAGCAAG CAGCCAACCA GGCAGGTTCT GTCCCTTTCA CTCACTGGCC CAAGGCGCCA 120
 CATCTCCCTC CAGAAAAGAC ACCATGAGCA CAGAAAGCAT GATCCGCGAC GTGGAAGTGG 180
 CAGAAGAGGC ACTCCCCCAA AAGATGGGGG GCTTCCAGAA CTCCAGGCGG TGCCTATGTC 240
 TCAGCCTCTT CTCATTCTG CTTGTGGCAG GGGCCACCAC GCTCTTCTGT CTAAGTGAAGT 300
 TCGGGGTGAT CGGTCCCCAA AGGGATGAGA AGTTCCCAA TGGCCTCCCT CTCATCAGTT 360

CTATGGCCCA GACCCTCACA CTCAGATCAT CTTCTCAAAA TTCGAGTGAC AAGCCTGTAG 420
 CCCACGTCGT AGCAAACCAC CAAGTGGAGG AGCAGCTGGA GTGGCTGAGC CAGCGCGCCA 480
 ACGCCCTCCT GGCCAACGGC ATGGATCTCA AAGACAACCA ACTAGTGGTG CCAGCCGATG 540
 GGTGTGACCT TGTCTACTCC CAGGTTCTCT TCAAGGGACA AGGCTGCCCC GACTACGTGC 600
 TCCTCACCCA CACCGTCAGC CGATTTGCTA TCTCATACCA GGAGAAAGTC AACCTCCTCT 660
 CTGCCGTCAA GAGCCCCTGC CCCAAGGACA CCCCTGAGGG GGCTGAGCTC AAACCCTGGT 720
 ATGAGCCCAT ATACCTGGGA GGAGTCTTCC AGCTGGAGAA GGGGGACCAA CTCAGCGCTG 780
 AGGTCAATCT GCCCAAGTAC TTAGACTTTG CGGAGTCCGG GCAGGTCTAC TTTGGAGTCA 840
 TTGCTCTGTG AAGGGAATGG GTGTTTCATCC ATTCTCTACC CAGCCCCCAC TCTGACCCCT 900
 TTA CTCTGAC CCCTTTATTG TCTACTCCTC AGAGCCCCCA GTCTGTGTCC TTCTAACTTA 960
 GAAAGGGGAT TATGGCTCAG AGTCCAATC TGTGCTCAGA GCTTTCAACA ACTACTCAGA 1020
 AACACAAGAT GCTGGGACAG TGACCTGGAC TGTGGGCCTC TCATGCACCA CCACCCACGG 1080
 AATCGAGAAA GAGCTATCAA TCTGGAATTC ACTGGAGCCT CGAATGTCCA TTCCTGAGTT 1140
 CTGCAAAGGG AGAGTGGTCA GGTTCCTCT GTCTCAGAAT GAGGCTGGAT AAGATCTCAG 1200
 GCCTTCCTAC CTTCAGACCT TTCCAGACTC TTCCCTGAGG TGCAATGCAC AGCCTTCCTC 1260
 ACAGAGCCAG CCCCCCTCTA TTTATATTTG CACTTATTAT TTATTATTTA TTTATTATTT 1320
 ATTTATTTGC TTATGAATGT ATTTATTTGG AAGGCCGGGG TGTCTGAGG GACCCAGTGT 1380
 GGGAAAGCTGT CTTCAGACAG ACATGTTTTT TGTGAAAACG GAGCTGAGCT GTCCCCACCT 1440
 GGCCTCTCTA CTTGTGTGCC TCCTCTTTTG CTTATGTTTA AAACAAAATA TTTATCTAAC 1500
 CCAATGTCT TAATAACGCT GATTTGGTGA CCAGGCTGTC GCTACATCAC TGAACCTCTG 1560
 CTCCCCACGG GAGCCGTGAC TGTAATTGCC CTACGGGTCA TTGAGAGAAA TAAAGATCGC 1620
 TTGGAAAAG 1629

(2) INFORMATION FOR SEQ ID NO:6:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 4110 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: Mus musculus

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

GAGACTCTGG	CCCCACGGGA	CACAGTGTC	CTGGTTTGAA	ACTTCTCAGC	CACCTTGGTG	60
AAGGGACTGA	GCTGTTAGAG	ACACTTCTGA	GGCTCCTCAC	GCTTGGGTCT	TGTTCACTCC	120
ACGGAGTAGC	CTAGTCAACT	GCAAGAGAAC	GGAGAACGTT	GGATTTGGAG	CAGAAGTGCA	180
AAGTCTCAGA	CATGGCTTGC	CCCTGGAAGT	TTCTCTTCAA	AGTCAAATCC	TACCAAAGTG	240
ACCTGAAAGA	GGAAAAGGAC	ATTAACAACA	ACGTGAAGAA	AACCCCTTGT	GCTGTTCTCA	300
GCCCAACAAT	ACAAGATGAC	CCTAAGAGTC	ACCAAAATGG	CTCCCCGCAG	CTCCTCACTG	360
GGACAGCACA	GAATGTTCCA	GAATCCCTGG	ACAAGCTGCA	TGTGACATCG	ACCCGTCCAC	420
AGTATGTGAG	GATCAAAAAC	TGGGGCAGTG	GAGAGATTTT	GCATGACACT	CTTCACCACA	480
AGGCCACATC	GGATTTCACT	TGCAAGTCCA	AGTCTTGCTT	GGGGTCCATC	ATGAACCCCA	540
AGAGTTTGAC	CAGAGGACCC	AGAGACAAGC	CTACCCCTCT	GGAGGAGCTC	CTGCCTCATG	600
CCATTGAGTT	CATCAACCAG	TATTATGGCT	CCTTTAAAGA	GGCAAAAATA	GAGGAACATC	660
TGGCCAGGCT	GGAAGCTGTA	ACAAAGGAAA	TAGAAACAAC	AGGAACCTAC	CAGCTCACTC	720
TGGATGAGCT	CATCTTTGCC	ACCAAGATGG	CCTGGAGGAA	TGTCCCTCGC	TGCATCGGCA	780
GGATCCAGTG	GTCCAACCTG	CAGGTCTTTG	ACGCTCGGAA	CTGTAGCACA	GCACAGGAAA	840
TGTTTCAGCA	CATCTGCAGA	CACATACTTT	ATGCCACCAA	CAATGGCAAC	ATCAGGTCGG	900
CCATCACTGT	GTTCCCCCAG	CGGAGTGACG	GCAAACATGA	CTTCAGGCTC	TGGAATTCAC	960
AGCTCATCCG	GTACGCTGGC	TACCAGATGC	CCGATGGCAC	CATCAGAGGG	GATGCTGCCA	1020
CCTTGGAGTT	CACCCAGTTG	TGCATCGACC	TAGGCTGGAA	GCCCCGCTAT	GGCCGCTTTG	1080
ATGTGCTGCC	TCTGGTCTTG	CAAGCTGATG	GTCAAGATCC	AGAGGTCTTT	GAAATCCCTC	1140
CTGATCTTGT	GTTGGAGGTG	ACCATGGAGC	ATCCCAAGTA	CGAGTGGTTC	CAGGAGCTCG	1200
GGTTGAAGTG	GTATGCACTG	CCTGCCGTGG	CCAACATGCT	ACTGGAGGTG	GGTGGCCTCG	1260
AATTCCCAGC	CTGCCCCTTC	AATGGTTGGT	ACATGGGCAC	CGAGATTGGA	GTTTCGAGACT	1320
TCTGTGACAC	ACAGCGCTAC	AACATCCTGG	AGGAAGTGGG	CCGAAGGATG	GGCCTGGAGA	1380
CCCACACACT	GGCCTCCCTC	TGGAAAGACC	GGGCTGTCAC	GGAGATCAAT	GTGGCTGTGC	1440
TCCATAGTTT	CCAGAAGCAG	AATGTGACCA	TCATGGACCA	CCACACAGCC	TCAGAGTCCT	1500
TCATGAAGCA	CATGCAGAAT	GAGTACCGGG	CCCGTGGAGG	CTGCCCCGCA	GACTGGATTT	1560
GGCTGGTCCC	TCCAGTGTCT	GGGAGCATCA	CCCCTGTGTT	CCACCAGGAG	ATGTTGAACT	1620
ATGTCCTATC	TCCATTCTAC	TACTACCAGA	TCGAGCCCTG	GAAGACCCAC	ATCTGGCAGA	1680
ATGAGAAGCT	GAGGCCCAGG	AGGAGAGAGA	TCCGATTTAG	AGTCTTGGTG	AAAGTGGTGT	1740
TCTTTGCTTC	CATGCTAATG	CGAAAGGTCA	TGGCTTCACG	GGTCAGAGCC	ACAGTCCTCT	1800

TTGCTACTGA	GACAGGGAAG	TCTGAAGCAC	TAGCCAGGGA	CCTGGCCACC	TTGTTCAGCT	1860
ACGCCTTCAA	CACCAAGGTT	GTCTGCATGG	ACCAGTATAA	GGCAAGCACC	TTGGAAGAGG	1920
AGCAACTACT	GCTGGTGGTG	ACAAGCACAT	TTGGGAATGG	AGACTGTCCC	AGCAATGGGC	1980
AGACTCTGAA	GAAATCTCTG	TTCATGCTTA	GAGAACTCAA	CCACACCTTC	AGGTATGCTG	2040
TGTTTGGCCT	TGGCTCCAGC	ATGTACCCTC	AGTTCTGCGC	CTTTGCTCAT	GACATCGACC	2100
AGAAGCTGTC	CCACCTGGGA	GCCTCTCAGC	TTGCCCCAAC	AGGAGAAGGG	GACGAACTCA	2160
GTGGGCAGGA	GGATGCCTTC	CGCAGCTGGG	CTGTACAAAC	CTTCCGGGCA	GCCTGTGAGA	2220
CCTTTGATGT	CCGAAGCAAA	CATCACATTC	AGATCCCGAA	ACGCTTCACT	TCCAATGCAA	2280
CATGGGAGCC	ACAGCAATAT	AGGCTCATCC	AGAGCCCGGA	GCCTTTAGAC	CTCAACAGAG	2340
CCCTCAGCAG	CATCCATGCA	AAGAACGTGT	TTACCATGAG	GCTGAAATCC	CAGCAGAATC	2400
TGCAGAGTGA	AAAGTCCAGC	CGCACCACCC	TCCTCGTTCA	GCTCACCTTC	GAGGGCAGCC	2460
GAGGGCCCAG	CTACCTGCCT	GGGGAACACC	TGGGGATCTT	CCCAGGCAAC	CAGACCGCCC	2520
TGGTGCAGGG	AATCTTGGAG	CGAGTTGTGG	ATTGTCCTAC	ACCACACCAA	ACTGTGTGCC	2580
TGGAGGTTCT	GGATGAGAGC	GGCAGCTACT	GGGTCAAAGA	CAAGAGGCTG	CCCCCCTGCT	2640
CACTCAGCCA	AGCCCTCACC	TACTTCCTGG	ACATTACGAC	CCCTCCCACC	CAGCTGCAGC	2700
TCCACAAGCT	GGCTCGCTTT	GGCACGGACG	AGACGGATAG	GCAGAGATTG	GAGGCCTTGT	2760
GTCAGCCCTC	AGAGTACAAT	GACTGGAAGT	TCAGCAACAA	CCCCACGTTC	CTGGAGGTGC	2820
TTGAAGAGTT	CCCTTCCTTG	CATGTGCCCC	CTGCCTTCCT	GCTGTCGCAG	CTCCCTATCT	2880
TGAAGCCCCG	CTACTACTCC	ATCAGCTCCT	CCCAGGACCA	CACCCCTCG	GAGGTTCCACC	2940
TCACTGTGGC	CGTGGTCACC	TACCGCACCC	GAGATGGTCA	GGGTCCCCTG	CACCATGGTG	3000
TCTGCAGCAC	TTGGATCAGG	AACCTGAAGC	CCCAGGACCC	AGTGCCCTGC	TTTGTGCGAA	3060
GTGTCAGTGG	CTTCCAGCTC	CCTGAGGACC	CCTCCCAGCC	TTGCATCCTC	ATTGGGCCCTG	3120
GTACGGGCAT	TGCTCCCTTC	CGAAGTTTCT	GGCAGCAGCG	GCTCCATGAC	TCCCAGCACA	3180
AAGGGCTCAA	AGGAGGCCGC	ATGAGCTTGG	TGTTTGGGTG	CCGGCACCCG	GAGGAGGACC	3240
ACCTCTATCA	GGAAGAAATG	CAGGAGATGG	TCCGCAAGAG	AGTGCTGTTC	CAGGTGCACA	3300
CAGGCTACTC	CCGGCTGCCC	GGCAAACCCA	AGGTCTACGT	TCAGGACATC	CTGCAAAAGC	3360
AGCTGGCCAA	TGAGGTACTC	AGCGTTCTCC	ACGGGGAGCA	GGGCCACCTC	TACATTTGCG	3420
GAGATGTGCG	CATGGCTCGG	GATGTGGCTA	CCACATTGAA	GAAGCTGGTG	GCCACCAAGC	3480
TGAACTTGAG	CGAGGAGCAG	GTGGAAGACT	ATTTCTTCCA	GCTCAAGAGC	CAGAAACGTT	3540
ATCATGAAGA	TATCTTCGGT	GCAGTCTTTT	CCTATGGGGC	AAAAAAGGGC	AGCGCCTTGG	3600
AGGAGCCCCA	AGCCACGAGG	CTCTGACAGC	CCAGAGTTCC	AGCTTCTGGC	ACTGAGTAAA	3660

GATAATGGTG AGGGGCTTGG GGAGACAGCG AAATGCAATC CCCCCCAAGC CCCTCATGTC 3720
 ATTCCCCCCT CCTCCACCCT ACCAAGTAGT ATTGTATTAT TGTGGACTAC TAAATCTCTC 3780
 TCCTCTCCTC CCTCCCCTCT CTCCCTTTCC TCCCTTCTTC TCCACTCCCC AGCTCCCTCC 3840
 TTCTCCTTCT CCTCCTTTGC CTCTCACTCT TCCTTGGAGC TGAGAGCAGA GAAAAACTCA 3900
 ACCTCCTGAC TGAAGCACTT TGGGTGACCA CCAGGAGGCA CCATGCCGCC GCTCTAATAC 3960
 TTAGCTGCAC TATGTACAGA TATTTATACT TCATATTTAA GAAAACAGAT ACTTTTGTCT 4020
 ACTCCCAATG ATGGCTTGGG CCTTTCTGT ATAATTCCTT GATGAAAAAT ATTTATATAA 4080
 AATACATTTT ATTTTAATCA AAAAAAAAAA 4110

(2) INFORMATION FOR SEQ ID NO:7:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 465 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (vi) ORIGINAL SOURCE:
 - (A) ORGANISM: *Rattus norvegicus*

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

GGCATCATGG CTGCCCTTCG GCCTCTGGTG AAGCCCAAGA TCGTCAAAAA GAGGACCAAG 60
 AAGTTCATCA GGCACCAGTC GGACCGATAT GTGAAAATTA AGCGAAACTG GCGGAAACCC 120
 AGAGGCATCG ACAACAGGGT GCGGAGAAGA TTCAAGGGCC AGATCCTGAT GCCCAACATT 180
 GGTACGGGA GTAACAAGAA AACCAAGCAC ATGCTGCCTA GCGGCTTCCG GAAGTTTCTG 240
 GTCCACAATG TCAAGGAGCT GGAAGTGCTG CTGATGTGCA ACAAATCTTA CTGTGCTGAG 300
 ATTGCTCACA ATGTGTCCTC TAAGAACCGA AAAGCCATCG TAGAAAGAGC AGCACAGCTG 360
 GCCATCAGAG TCACCAATCC CAACGCCAGG CTACGCAGCG AAGAGAATGA ATAGATGGCT 420
 TGTGTGCCTG TTTTGTGTTT AAATAAAACC ACAAAAACCTG CCAA 465

(2) INFORMATION FOR SEQ ID NO:8:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 21 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:

(A) ORGANISM: Mus musculus

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

GCTATCGACA AGCAGCAGAA T

21

(2) INFORMATION FOR SEQ ID NO:9:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 22 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:

(A) ORGANISM: Mus musculus

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

TGAACACAAC AACATAAAGC CC

22

(2) INFORMATION FOR SEQ ID NO:10:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 26 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:

(A) ORGANISM: Mus musculus

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

TGTTATATCT GGTTATTATT GAATGG

26

(2) INFORMATION FOR SEQ ID NO:11:

(i) SEQUENCE CHARACTERISTICS:

WE CLAIM:

1. A method for treating a patient having a condition in which regulating energy metabolism during a systemic inflammatory response is desired,
5 comprising administering a composition having a physiologically effective amount of at least one OB-R agonist ligand.
2. The method of claim 1 wherein the OB-R agonist ligand is recombinant human OB protein.
3. The method of claim 2 wherein the amount of recombinant
10 human OB protein administered is from about 1 microgram per kilogram body weight to about 50 micrograms per kilogram body weight.
4. The method of claim 1 wherein the OB-R agonist ligand is a peptide conformational analog of human OB protein comprising conservative substitutions of amino acid residues.
- 15 5. The method of claim 1 wherein the OB-R agonist ligand is an OB-related peptide.
6. The method of claim 1 wherein the condition is sepsis.
7. The method of claim 1 wherein the condition is systemic inflammatory response syndrome.
- 20 8. A composition for treating a patient having a condition in which regulating energy metabolism during a systemic inflammatory response is desired, comprising a physiologically effective amount of at least one OB-R agonist ligand.
9. The composition of claim 8 wherein the OB-R agonist
25 ligand is recombinant human OB protein.
10. The composition of claim 9 wherein the amount of recombinant human OB protein per dose is from about 1 microgram per kilogram body weight to about 50 micrograms per kilogram body weight.

11. The composition of claim 8 wherein the OB-R agonist ligand is a peptide conformational analog of human OB protein comprising conservative substitutions of amino acid residues.
12. The composition of claim 8 wherein the OB-R agonist
5 ligand is an OB-related peptide.
13. The composition of claim 8 wherein the condition is sepsis.
14. The composition of claim 8 wherein the condition is systemic inflammatory response syndrome.
15. A composition for the amelioration of the toxicity of
10 therapeutic cytokines comprising a physiologically effective amount of an OB-R agonist ligand.
16. The composition of claim 15 wherein the OB-R agonist ligand is recombinant human OB protein.
17. The composition of claim 15 wherein the amount of
15 recombinant human OB protein per dose is 1 microgram per kilogram body weight to about 50 micrograms per kilogram body weight.
18. A method for the treatment of a patient having obesity comprising the steps of:
- 20 administering at least one OB-R expression inducer; and
administering a physiologically effective amount of an OB-R agonist ligand.
19. The method of claim 18 wherein the OB-R expression inducer is a compound chosen from the group consisting of LPS, IL-1 α , IL-1 β , TNF- α and IL-6.
20. The method of claim 18 wherein the OB-R expression inducer and the OB-R agonist ligand are administered at a different times.
21. The method of claim 18 wherein the OB-R expression inducer is administered in an amount from about 0.003 to about 20 micrograms per kilogram body weight.

22. The method of claim 18 wherein the OB-R agonist ligand is administered in an amount from about 1 microgram per kilogram body weight to about 50 micrograms per kilogram body weight.

23. The method of claim 18 wherein the OB-R agonist ligand is recombinant human OB protein.

24. The method of claim 23 wherein the recombinant human OB protein is administered in an amount from about 1 micrograms per kilogram body weight to about 50 micrograms per kilogram body weight.

25. The method of claim 18 wherein the OB-R expression inducer is IL-6.

26. The method of claim 25 wherein IL-6 is administered in an amount from about 0.5 to about 20 micrograms per kilogram body weight.

27. A method for the treatment of a patient having a condition characterized by OB resistance, comprising the steps of:

administering IL-6 in an amount from about 0.5 to about 20 micrograms per kilogram body weight; and

administering recombinant human OB protein in an amount from about 1 microgram per kilogram body weight to about 50 micrograms per kilogram body weight.

28. A composition suitable for the treatment of obesity comprising:

at least one therapeutic cytokine capable of increasing the expression of the OB receptor;

a physiologically effective amount of an OB-R agonist ligand; and a pharmaceutically acceptable excipient.

29. The composition of claim 28 wherein the therapeutic cytokine capable of increasing the expression of the OB receptor and the OB-R agonist ligand are packaged separately.

30. The composition of claim 28 wherein the therapeutic cytokine is about 0.5 to about 20 micrograms per kilogram body weight IL-6.

31. The composition of claim 29 wherein the OB-R agonist ligand is administered in a dose of about 1 micrograms per kilogram body weight to about 50 micrograms per kilogram body weight recombinant human OB protein.

5 32. An assay kit for a disease marker in a sample for a systemic inflammatory response in a patient comprising:

an antibody capable of binding to OB protein; and

a detection means for determining the amount of the antibody bound to OB protein.

10 33. A method for assaying a disease marker for an inflammatory response in a patient comprising:

mixing a portion of the sample with an antibody capable of binding to OB protein; and

detecting the amount of antibody bound to OB protein.

15 34. A composition suitable for the treatment of anorexia, cachexia or other wasting condition comprising a physiologically effective amount of antibody capable of binding OB protein.

20 35. The method for the treatment of anorexia, cachexia or other wasting condition comprising administering a physiologically effective amount of antibody capable of binding OB protein in an amount from about 0.02 to about 15 milligrams per kilogram body weight per day.

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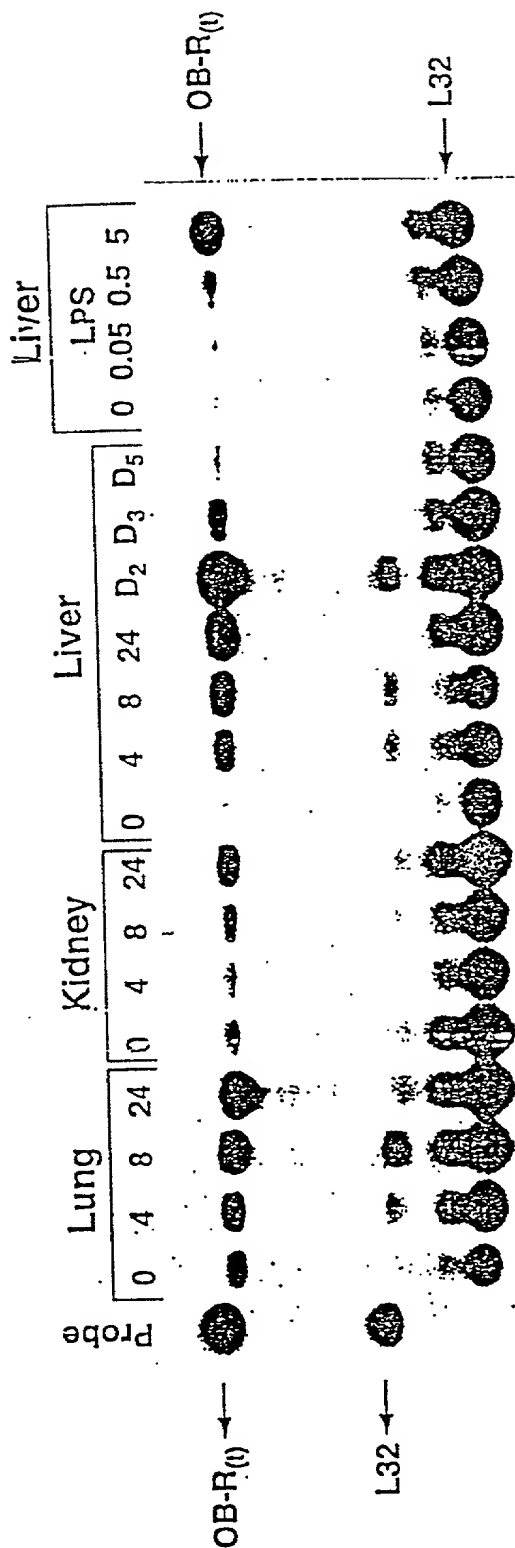


FIGURE 1

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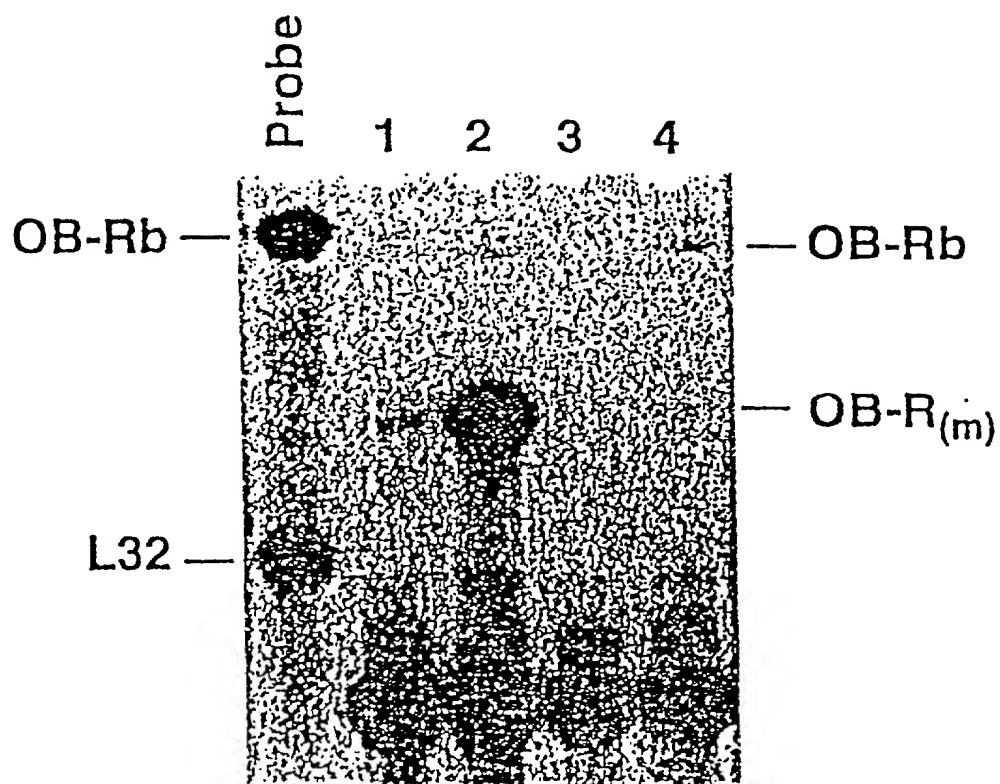


FIGURE 2

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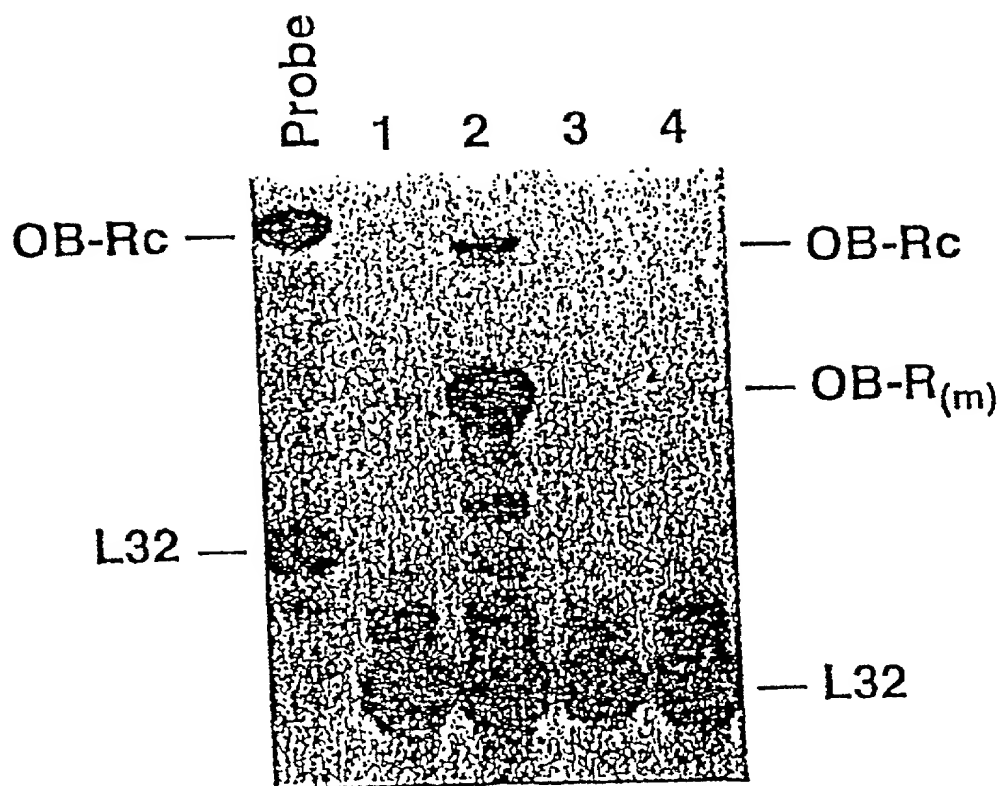


FIGURE 3

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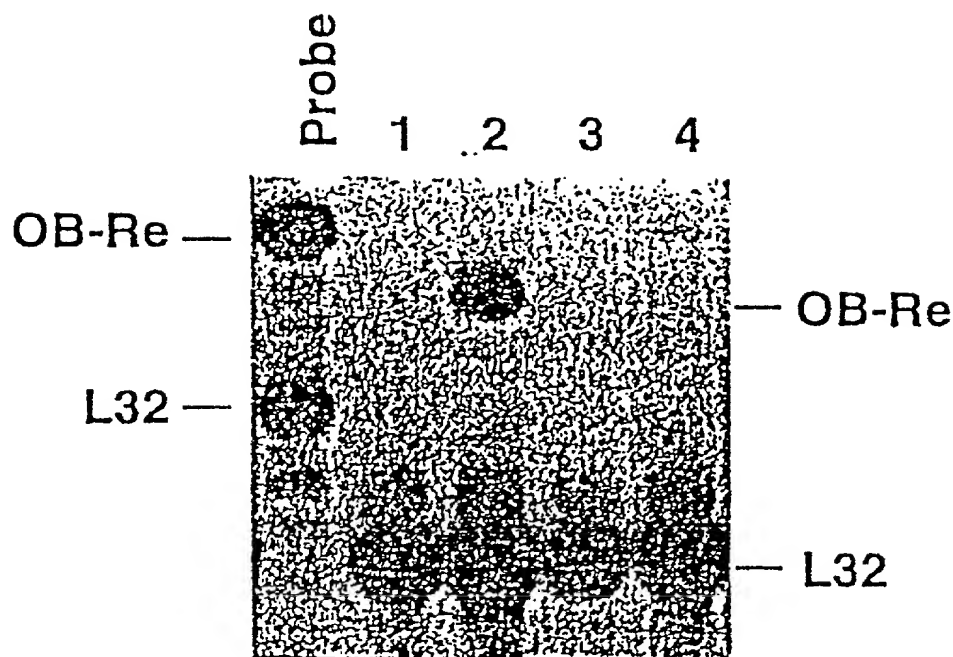


FIGURE 4

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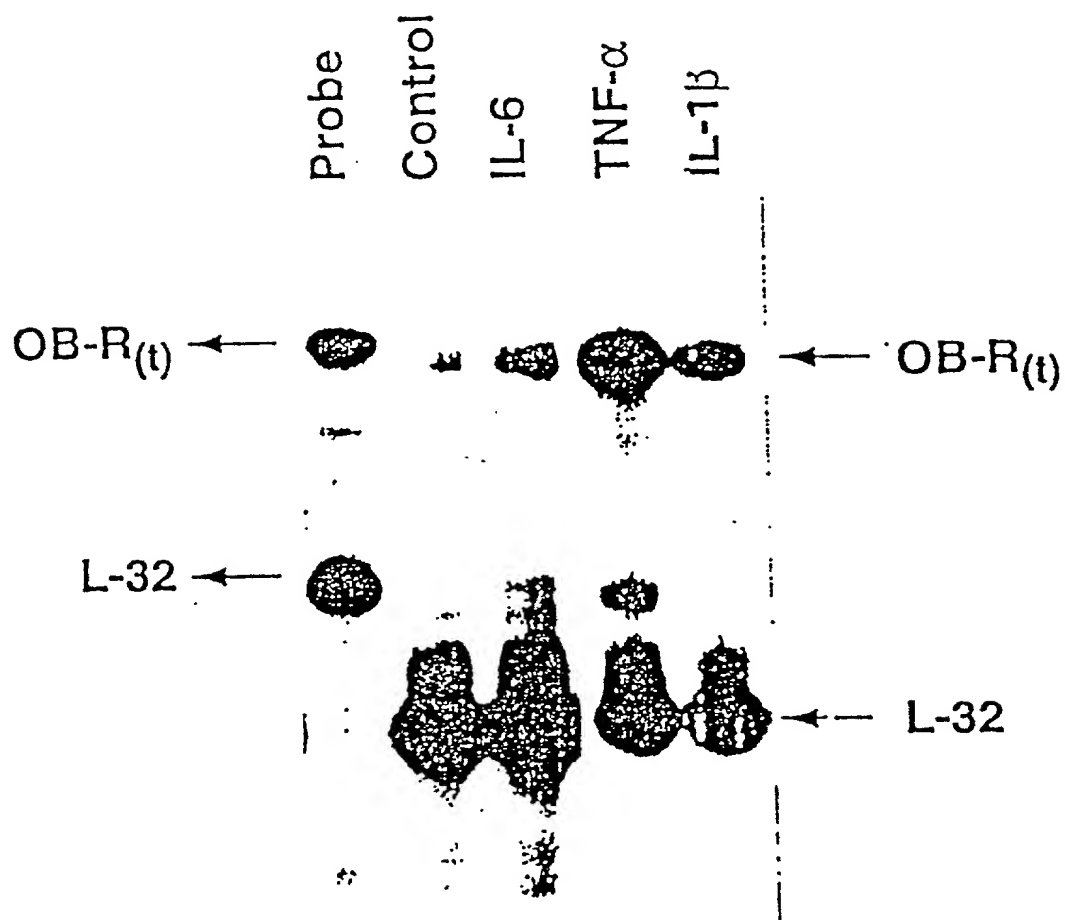


FIGURE 5

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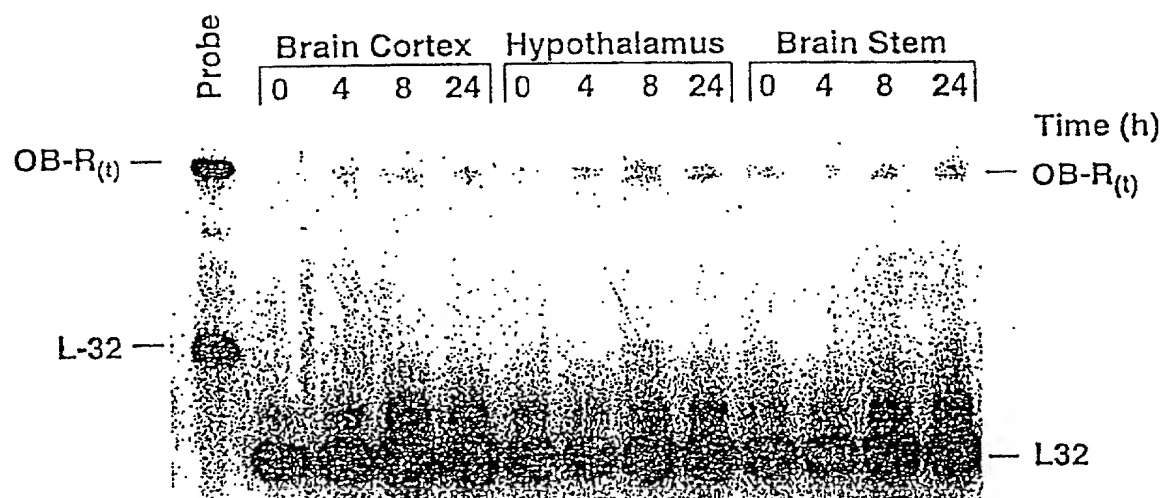


FIGURE 6

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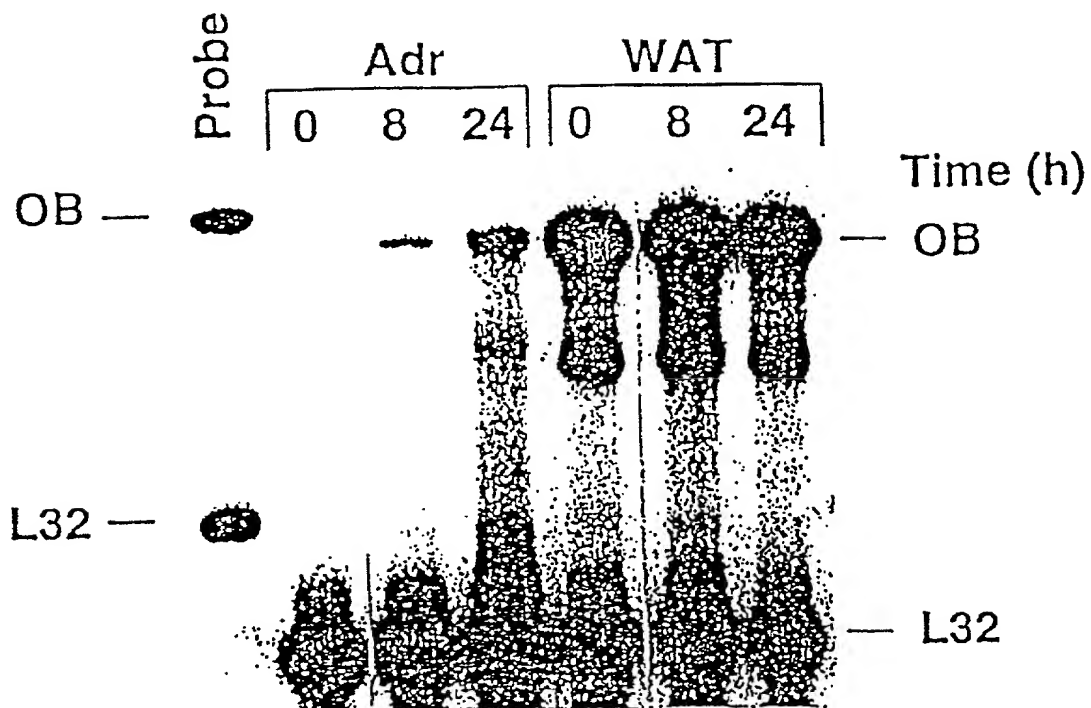


FIGURE 7

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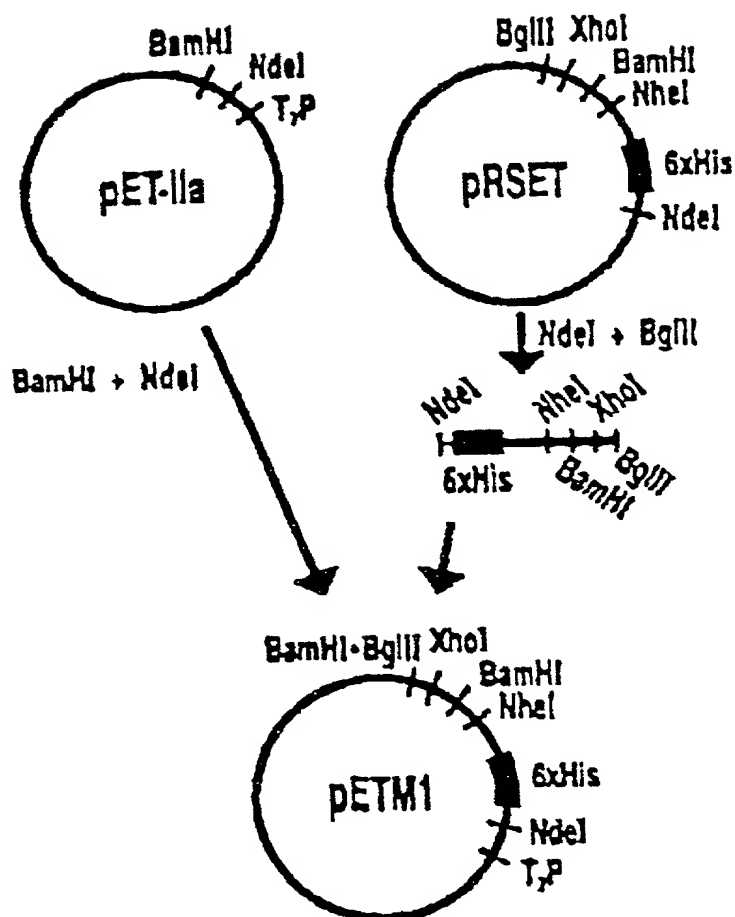


FIGURE 8

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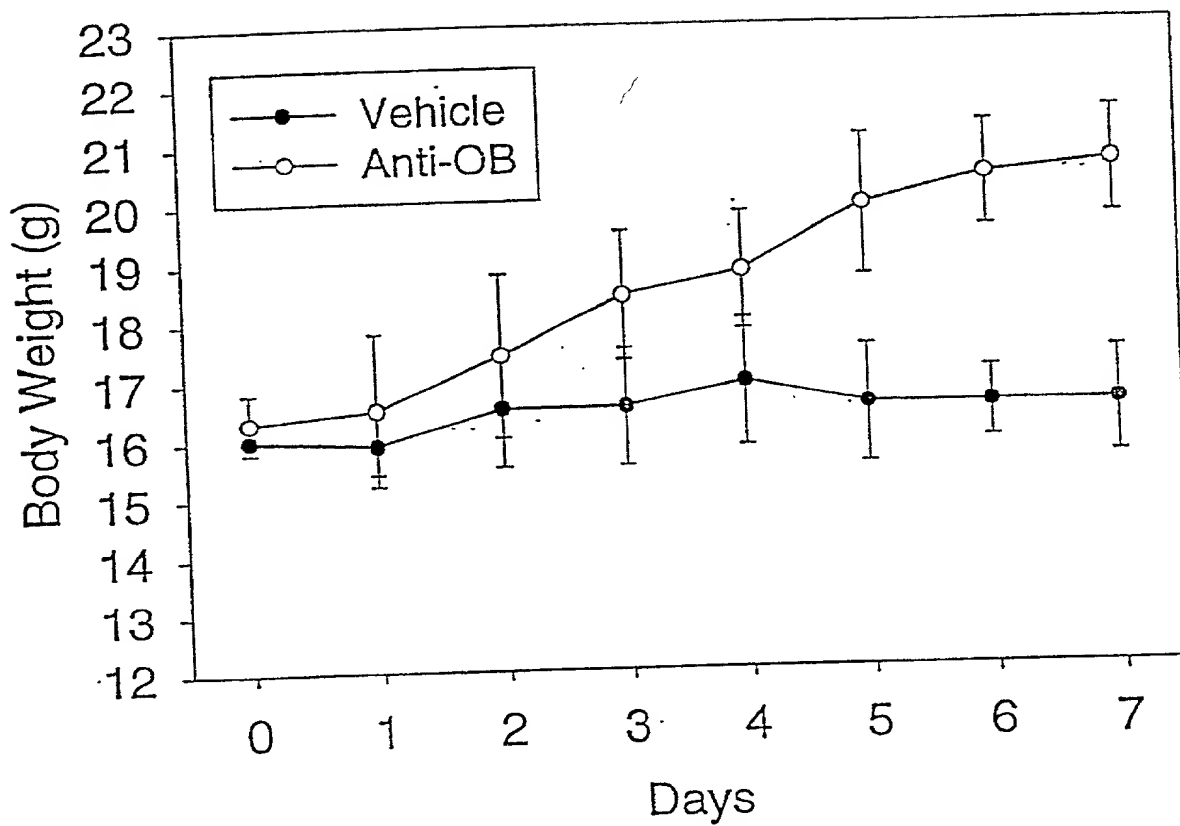


FIGURE 9

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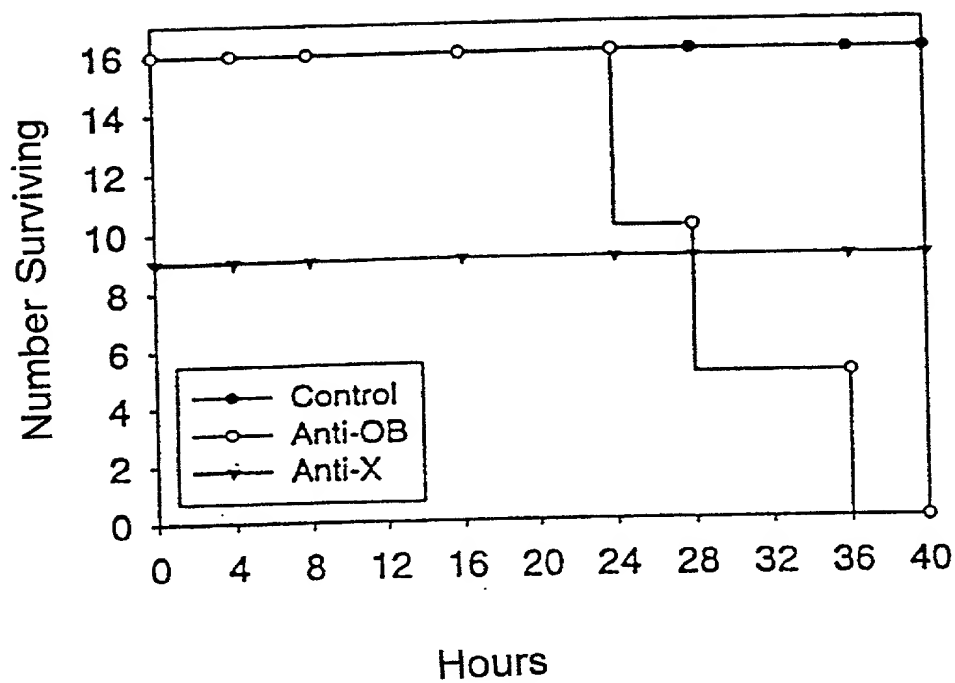


FIGURE 10

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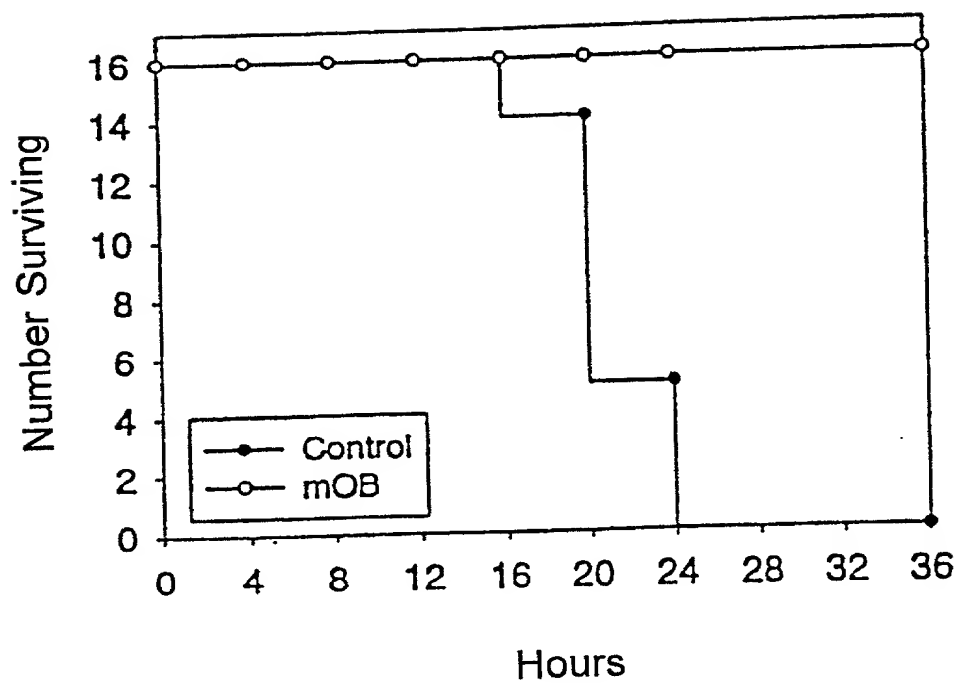


FIGURE 11

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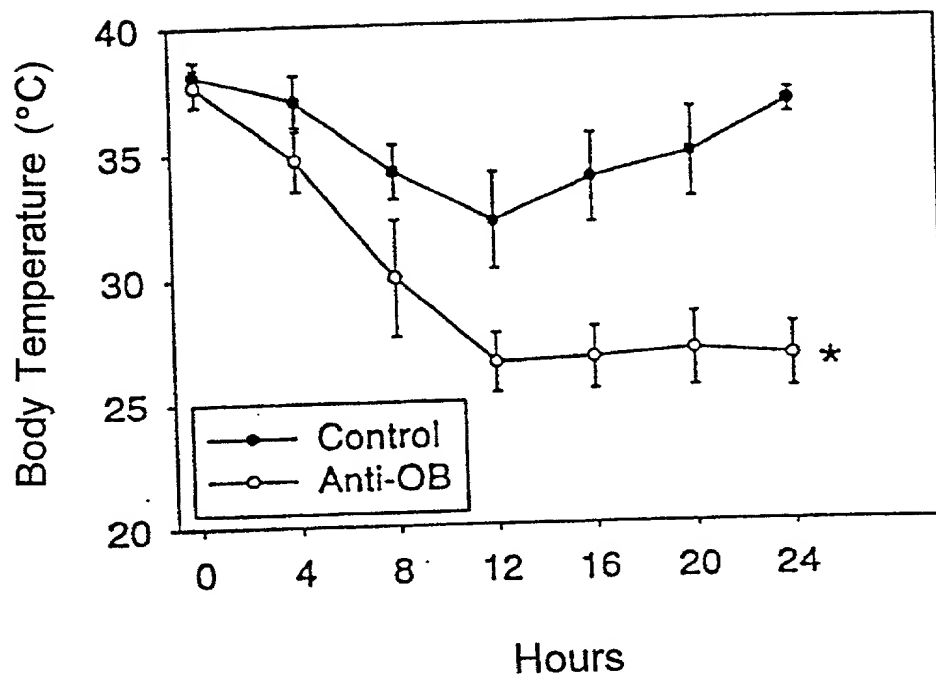


FIGURE 12

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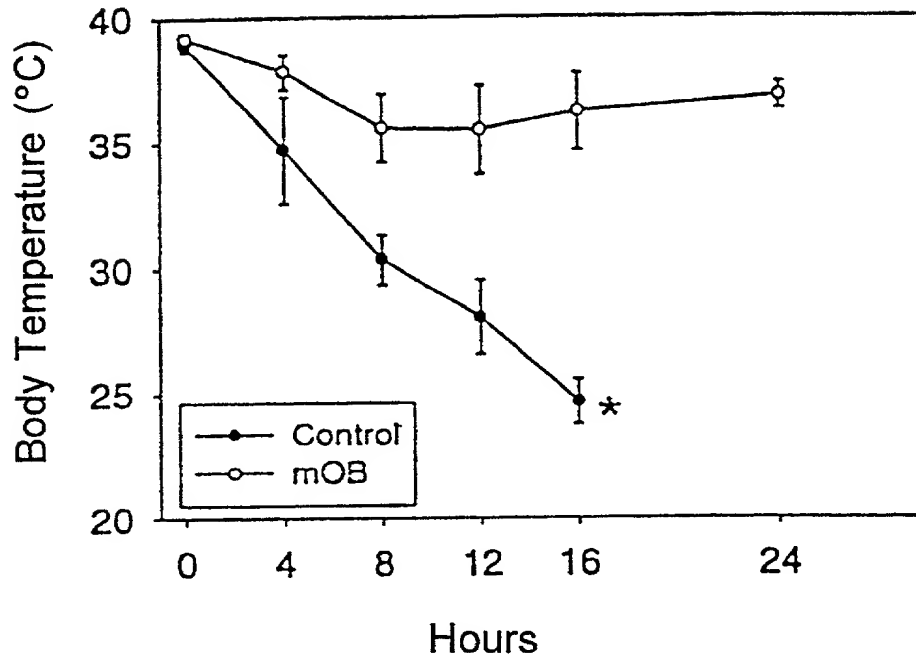


FIGURE 13

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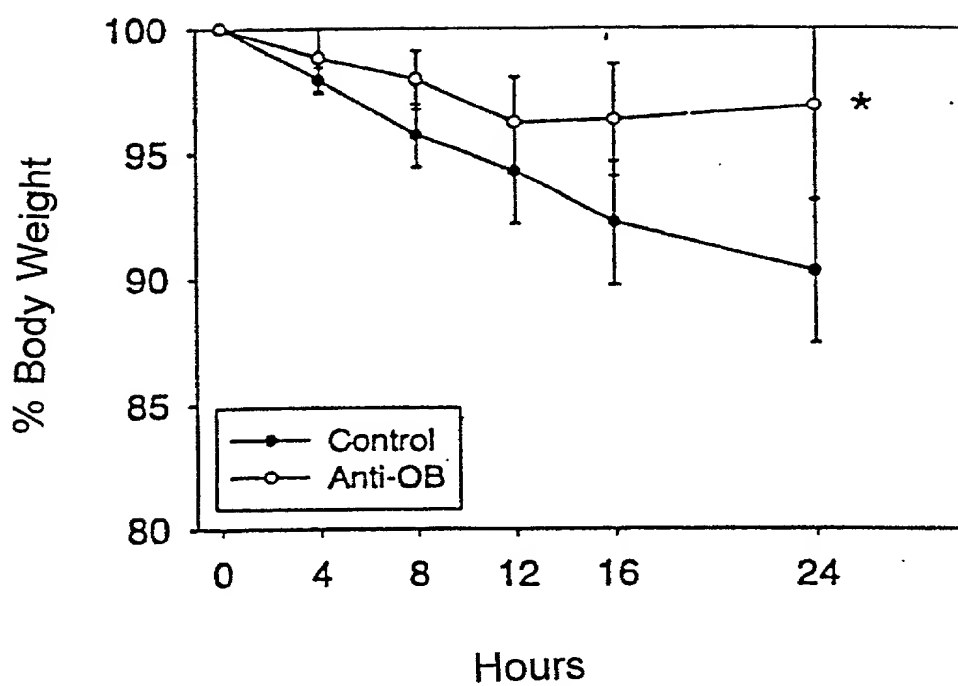
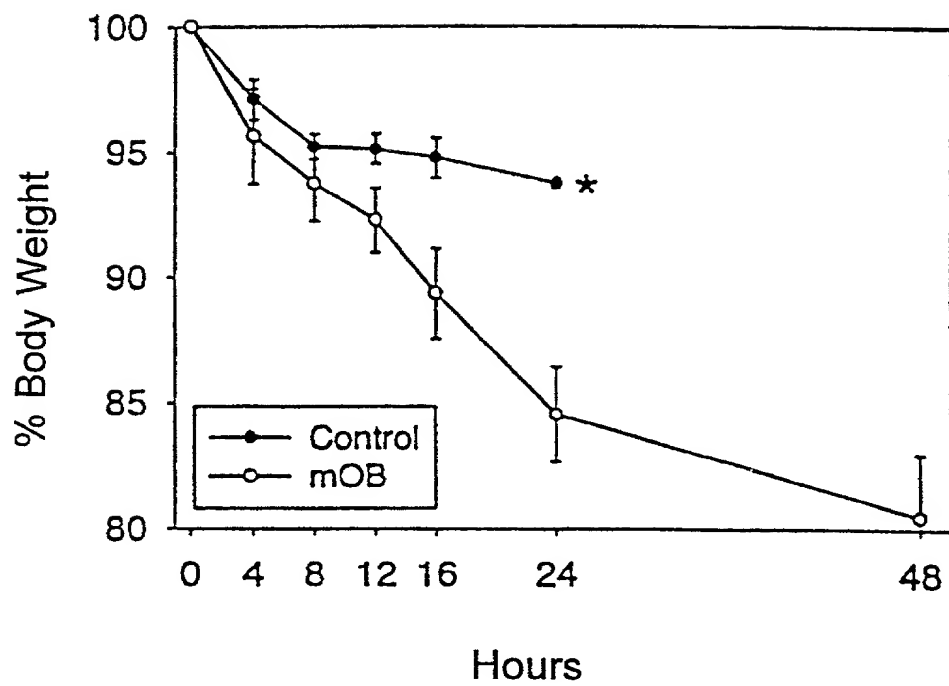


FIGURE 14

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**FIGURE 15**

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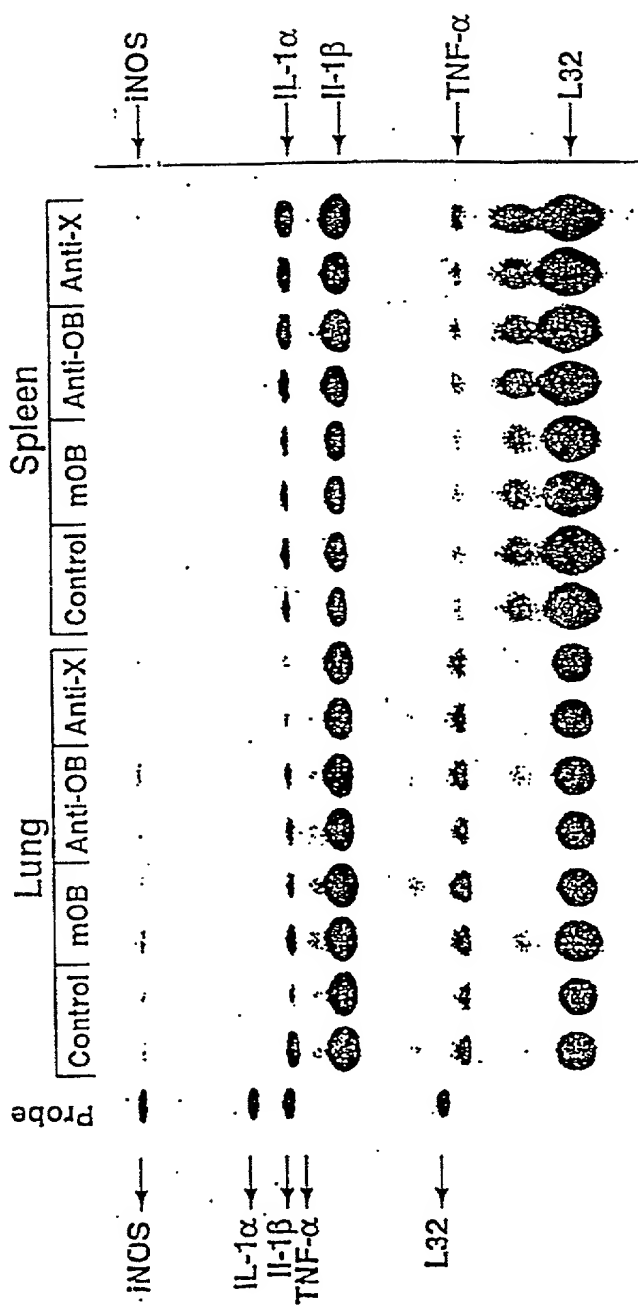


FIGURE 16

PATENT APPLICATION DECLARATION AND POWER OF ATTORNEY

I HEREBY DECLARE THAT:

My residence, post office address, and citizenship are as stated below.

I believe I am the original, first, and sole inventor (if only one name is listed) or an original, first, and joint inventor (if plural names are listed) of the subject matter which is claimed and for which a patent is sought on the invention entitled DIAGNOSTIC AND THERAPEUTIC METHODS RELATED TO REGULATING ENERGY MOBILIZATION WITH OB PROTEIN AND OB ANTIBODIES, the specification of which:

— is attached hereto;

X was filed on June 4, 1997 as Application Serial No. PCT/US97/09684
and was amended on N/A (if applicable).

I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment referred to herein.

I acknowledge the duty to disclose information to the Patent and Trademark Office known to me to be material to the patentability of this application, as defined in Title 37, Code of Federal Regulations, Sec. 1.56.

I do hereby declare that all statements made herein of my knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

I hereby appoint the following as my attorneys or agents with full power of substitution to prosecute this application and transact all business in the United States Patent and Trademark Office connected therewith:

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Dolores T. Kenney	Reg. No. 31,269	Talivaldis Cepuritis	Reg. No. 20,818
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Inventor's signature: [Signature] Date: 8/13/97

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Second Inventor's signature: [Signature] Date: 8/13/97

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Post Office Address (If different) _____

Third Inventor's signature: [Signature] Date: 8/13/97